

ANALYTICAL CHALLENGES IN DEVELOPING STRATEGIES FOR MYCOTOXIN PREVENTION AND CONTROL - ADVANCED AND RAPID METHODS FOR MULTI-TOXIN AND MULTI-BIOMARKER ANALYSIS

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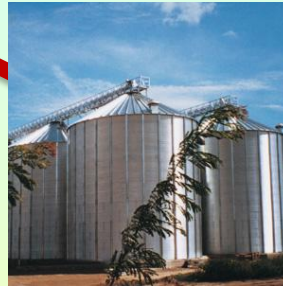
FUNGAL DEVELOPMENT AND MYCOTOXIN PRODUCTION IN FOOD



planting



harvest



storage

logistic



logistic

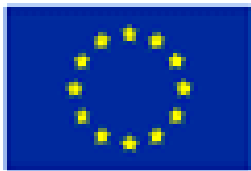


processing

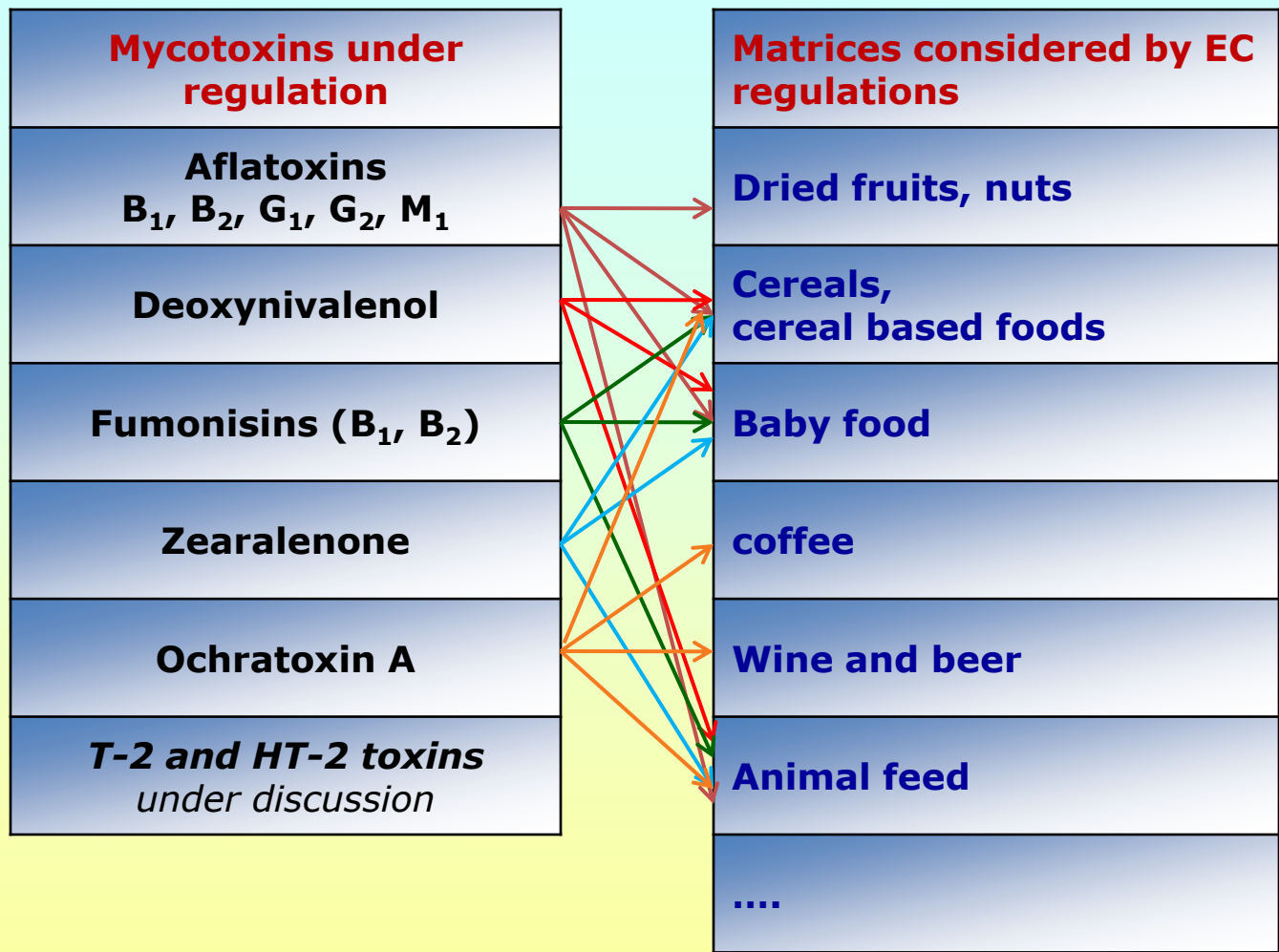
distribution



Need to improve **mycotoxin monitoring and **prevention** to minimize contamination at different critical steps of the food chain "from farm to fork"**



EC regulations 1881/2006 and 1126/2007 Maximum permitted levels of mycotoxins



Maximum levels range from:
ng/kg to **mg/kg**

Need of **reliable** analytical methods **applicable at regulatory levels** in a **wide range** of matrices

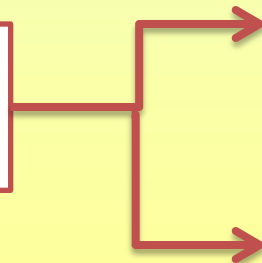
PRESENTATION OUTLINE

- ✓ **Multi-mycotoxin determination in food by LC-MS(MS)**
Tandem MS and high resolution MS approaches
- ✓ **Multi-biomarker determination in human urine by LC-MS/MS**
- ✓ **Multi-mycotoxin determination in food/feed by multiplex dipstick immunoassay**

**Multi-mycotoxin
analysis**

**Advanced
LC-MS(MS)**

**Dipstick for
rapid screening**



Challenges in multi-mycotoxin method development for food matrices

- ⇒ **Sample preparation**
- ⇒ **Analyte detection**
- ⇒ **Method validation**
- ⇒ **Wide range of applicability**

HPLC
coupled with
Mass Spectrometry

Determination of aflatoxins, ochratoxin A and *Fusarium* toxins in cereal-based products by LC-MS/MS or LC-HRMS after SPE clean up



Wheat flour

Barley flour

Oat flour



Wheat based crisp bread
Rye based crisp bread

Breakfast cereals
Cereal biscuits



Sample



Extraction

acetonitrile/water
(84/16)



Filtration (Whatmann N°4)

Evaporation

Reconstitution with methanol/water (10/90)

Clean-up with

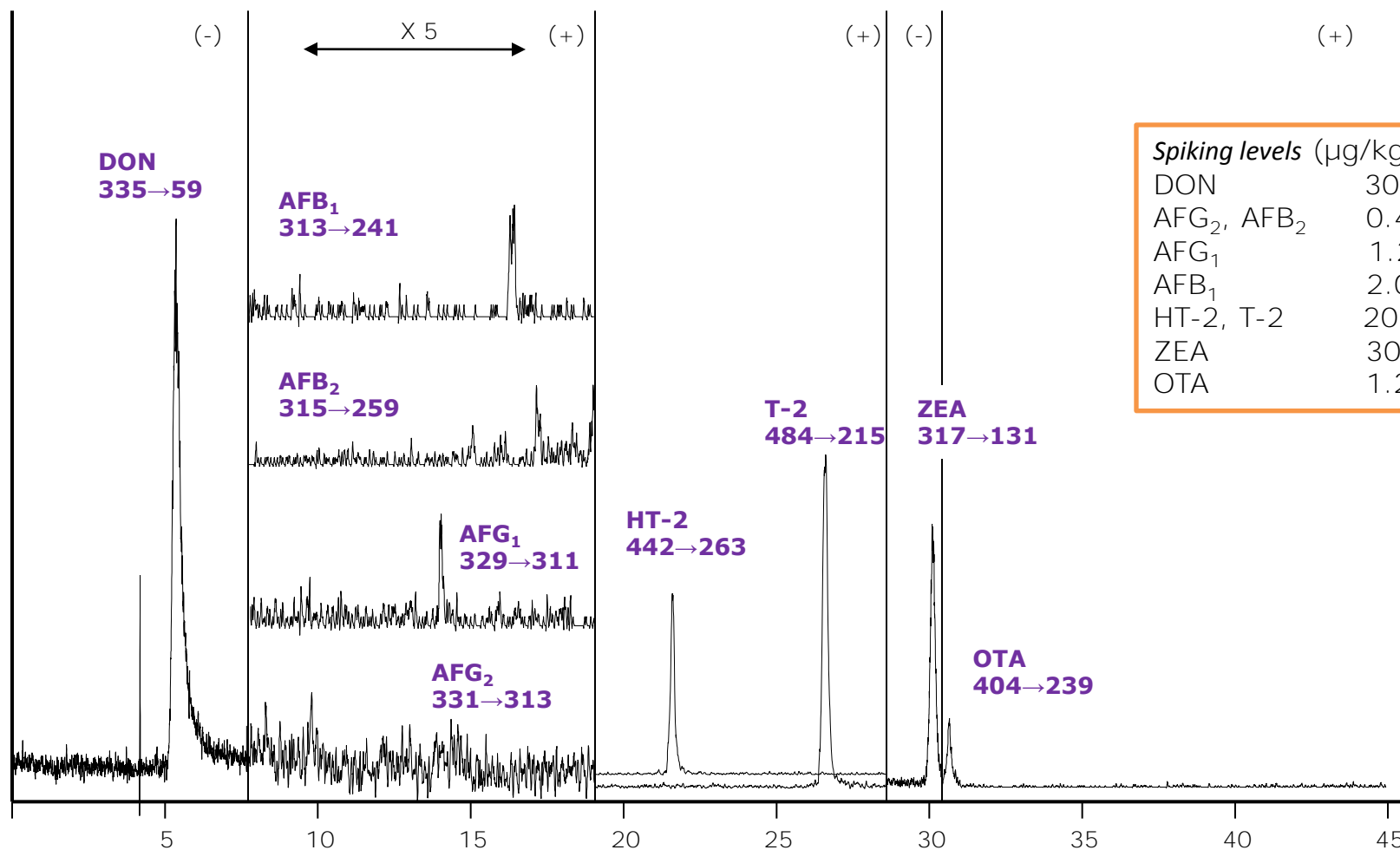
OASIS® HLB

(polimeric SPE cartridges)

LC-MS/MS analysis

LC-HRMS analysis

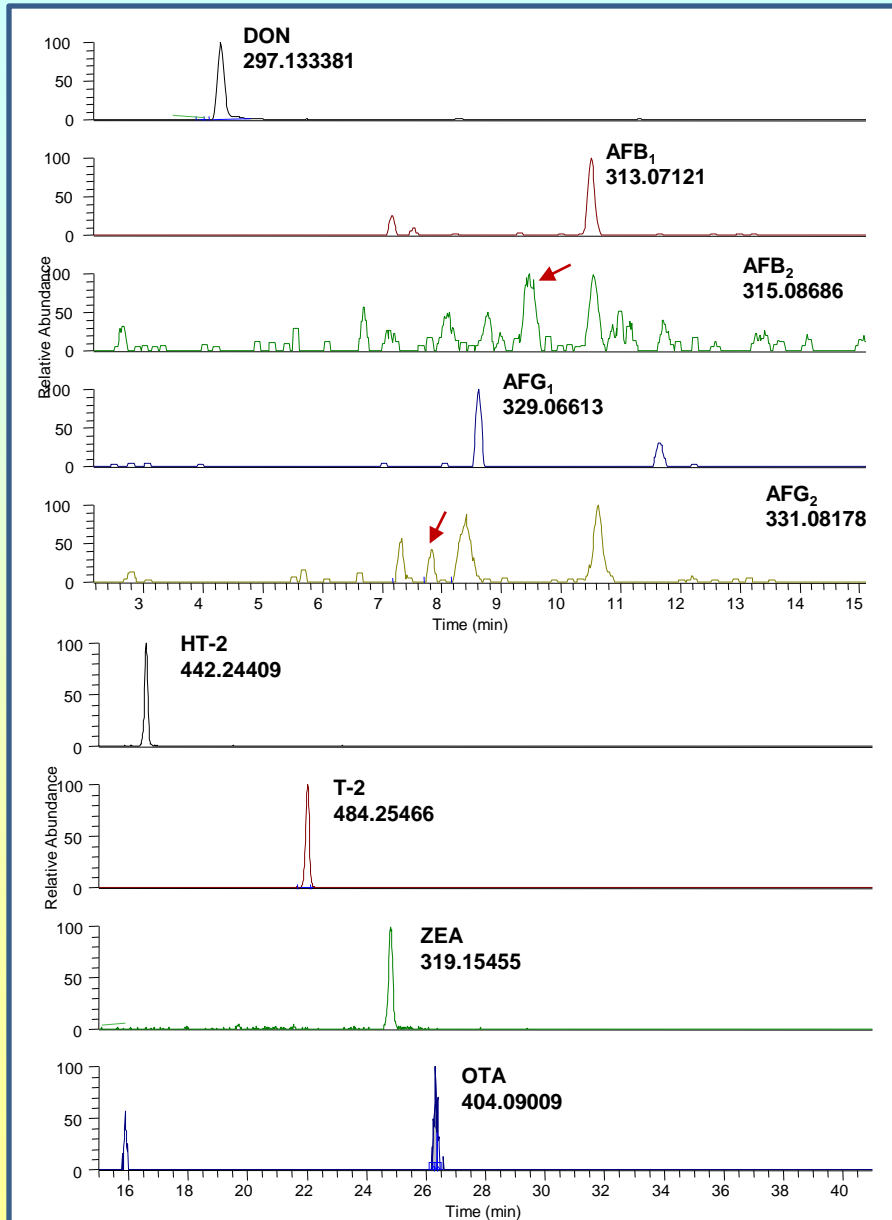
LC-MS/MS chromatogram of a spiked crisp bread extract



Spiking levels (µg/kg)	
DON	300
AFG ₂ , AFB ₂	0.4
AFG ₁	1.2
AFB ₁	2.0
HT-2, T-2	20
ZEA	30
OTA	1.2

Column: Gemini RP18 (150 × 2.0 mm, 5 µm) Phenomenex
Flow: 200 µl/min
Column oven: 40 °C
Solv A: H₂O, 0.5% acetic acid, 1mM AcNH₄
Solv B: CH₃OH, 0.5% acetic acid, 1mM AcNH₄
Injection volume: 20µl (100 mg sample)

LC-HRMS chromatogram of a spiked crisp bread extract



Spiking levels (µg/kg)

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Methods performances and comparison between LC-MS/MS and LC-HRMS detection

RECOVERIES and REPEATABILITY

EC acceptance criteria (401/2006)

Recoveries, % (RSDr %) in WHEAT BASED CRISP BREAD

Spiking level (µg/kg)	300	2	0.5	1.2	0.5	20	20	30	1.2
	DON	AFG₂	AFG₁	AFB₂	AFB₁	HT-2	T-2	ZEA	OTA
MS/MS	100 (0)	101 (6)	106 (5)	85 (10)	102 (6)	107 (2)	108 (6)	84 (5)	101 (3)
HRMS	104 (0)	102 (5)	104 (4)	80 (2)	102 (2)	105 (1)	103 (1)	85 (1)	93 (2)

*Similar results in:
barley, wheat and oat flours, rye-based crisp bread*

Methods performances and comparison between LC-MS/MS and LC-HRMS detection

DETECTION LIMITS

*EC maximum permitted levels
(EC regulations 1881/2006 and 1126/2007)*

LOD ($\mu\text{g}/\text{kg}$)		
	LC-HRMS	LC-MS/MS
DON	0.3 (+)	29.0 (-)
AFG₂	0.1	0.5
AFG₁	0.2	0.7
AFB₂	0.1	0.4
AFB₁	0.1	0.5
HT-2	0.3	0.5
T-2	0.3	0.5
ZEA	0.4	2.2
OTA	0.2	0.1

Methods performances and comparison between LC-MS/MS and LC-HRMS detection

MATRIX EFFECTS

Matrix effect SSE %		
	LC-HRMS	LC-MS/MS
DON	76	95
AFG₂	100	95
AFG₁	100	89
AFB₂	67	88
AFB₁	100	88
HT-2	90	95
T-2	94	103
ZEA	65	70
OTA	87	106

SSE = signal suppression/enhancement
= (slope of matrix calibration/slope of standard calibration)*100

When **robust sample preparation** and **good chromatographic separation** are applied, **similar matrix effects** are obtained with different MS instrumentation and detection modes.

Methods performances and comparison between LC-MS/MS and LC-HRMS detection

MS/MS detection HRMS detection legislation requirements

*EC performance criteria (2002/657/EC)
Doc No Sanco/10684/2009*

MS/MS	HRMS*
1 precursor ion 2 daughters ions	2 ions Mass accuracy < 5ppm
<i>example: AFB₁</i>	
313.0 – 241.1	313.07066
313.0 – 213.4	241.04953

* Fragment ion obtained by collision cell induced fragmentation (HCD)

Proficiency test for multi-mycotoxin methods based on LC-MS(MS)

***Aim of the study:* to obtain information on currently used LC-MS(MS) methodologies for multi-mycotoxin analysis and relevant performances.**

- ✓ **56 laboratories involved**
- ✓ **Distribution of materials ongoing (by Dec 15th, 2010)**
- ✓ **Results expected by February 2011.**

LC-MS/MS DETERMINATION OF MULTI-MYCOTOXIN BIOMARKER IN HUMAN AND ANIMAL URINE

- ✓ A reliable indication of individual exposure to the major mycotoxins may be provided by a biomarker
- ✓ Potential markers include the parent compound or metabolite

Mycotoxin	Urinary biomarker
aflatoxin B ₁	aflatoxin M ₁ (AFM ₁)
ochratoxin A	ochratoxin A (OTA)
deoxynivalenol	deoxynivalenol (DON)
	de-epoxydeoxynivalenol (DOM-1)
zearalenone	alfa-zearalenol (α-ZOL)
	beta-zearalenol (β-ZOL)
fumonisin B ₁	fumonisin B ₁ (FB ₁)

- ✓ The response of DON, DOM-1, AFM₁, FB₁, α-ZOL, β-ZOL, OTA as biomarker of mycotoxin exposure has been demonstrated in pig, rat and mouse.

Challenges in multi-mycotoxin method development for biological fluids

Lack of suitable methods of analysis to detect simultaneously a range of chemically different metabolites at trace levels in biological fluids

⇒ **Sensitivity** (up to pg/ml)

⇒ **Sample preparation**

⇒ **Analyte detection**

⇒ **Method validation**

HPLC
coupled with
Mass Spectrometry

Sample preparation and analysis

Urine sample
Dilution with water



MYCO6in1
LC/MS/MS

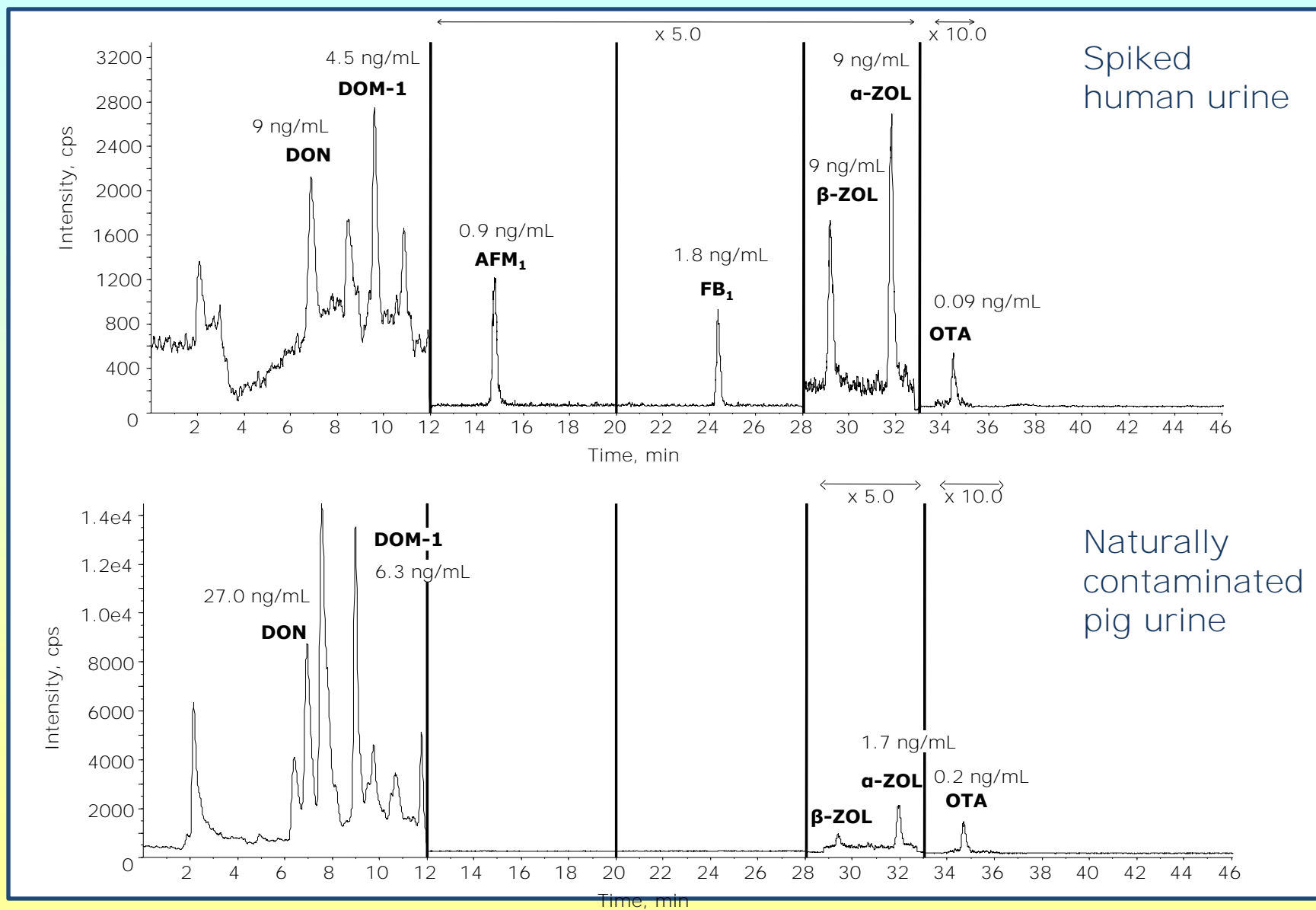


Clean up
Immunoaffinity column + OASIS HLB column

LC-MS/MS analysis

DON
DOM-1
AFM₁
FB₁
α-ZOL, β-ZOL
OTA

MRM chromatograms of urine samples after IAC-SPE clean up



METHOD PERFORMANCES

Mean Recoveries, % (RSDr %) in Human Urine

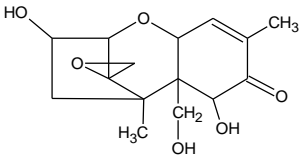
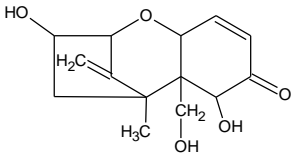
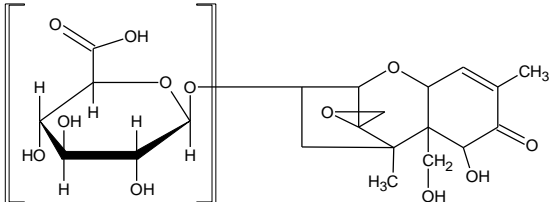
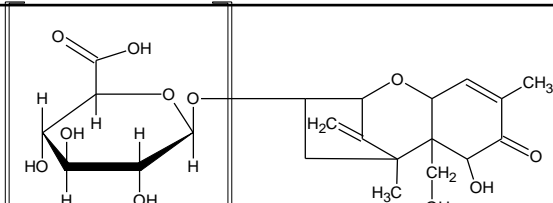
Spiking level range(ng/ml)	3-12			1.5-6.0	0.3-1.2	0.6-2.4	0.03-0.12
Mycotoxin	DON	α-ZOL	β-ZOL	DOM-1	AFM₁	FB₁	OTA
Recovery (RSDr)	77 (13)	72 (9)	83 (18)	78 (9)	96 (8)	62 (3)	65 (8)

Limits of Detection (S/N = 3) in Human Urine (ng/ml)

DON	α-ZOL	β-ZOL	DOM-1	AFM₁	FB₁	OTA
0.8	0.8	2.2	0.8	0.06	0.1	0.02

LC-MS/MS CHARACTERIZATION OF DON URINARY METABOLITE PROFILE IN HUMAN AND RATS

V.M.T. Lattanzio, M. Solfrizzo, A. De Girolamo, S. Chulze, A. Torres, A. Visconti, J. Chrom. B. XXX

	Molecular structure	Characteristic ions (m/z)	Presence in urine	
			Rat	Human
DON		355.1 295.0 265.1 59.0 <i>Negative ions</i>	YES	YES
DOM-1		339.1 279.0 249.1 59.0 <i>Negative ions</i>	YES	NO
DON glucuronide1		490.2 297.2 249.2 231.1 <i>Positive ions</i>	YES	YES
DON glucuronide2		490.2 177.1 103.1 89.0 <i>Positive ions</i>	NO	YES
DOM-1 glucuronide		474.2 281.2 233.1 130.1 <i>Positive ions</i>	YES	YES

the position of glucuronide moiety is indicative

Challenges in multi-mycotoxin method development

Rapid methods based on dipstick immunoassays

⇒ **Antibody production and characterization**

⇒ **Multiplex dipstick design and assembly**

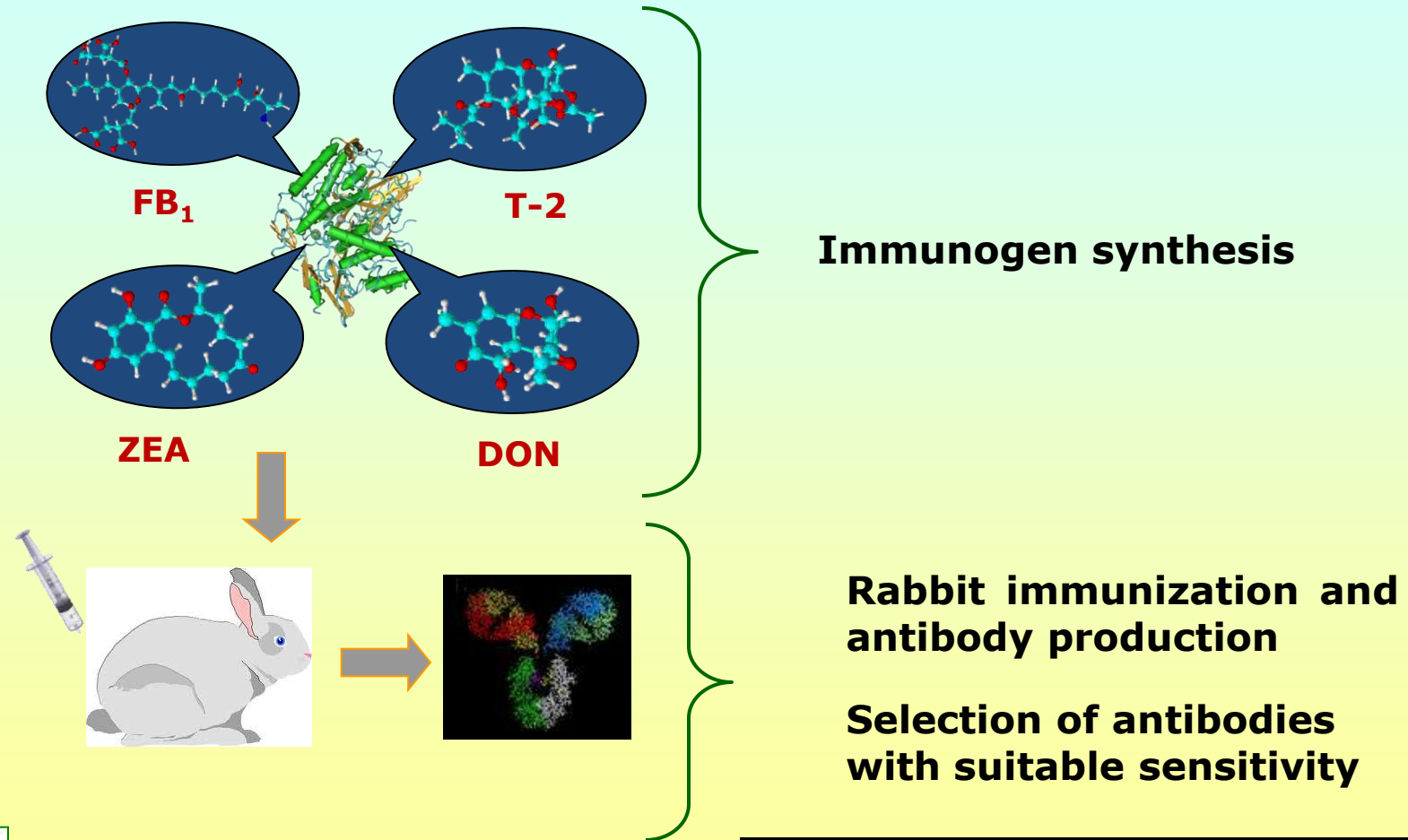
⇒ **Development of simplified sample preparation protocols**

⇒ **Method validation and application**



Antibody production and characterization

Polyclonal antibodies raised against **T-2/HT-2, DON, ZEA** and **FB₁/FB₂** have been produced on rabbits.

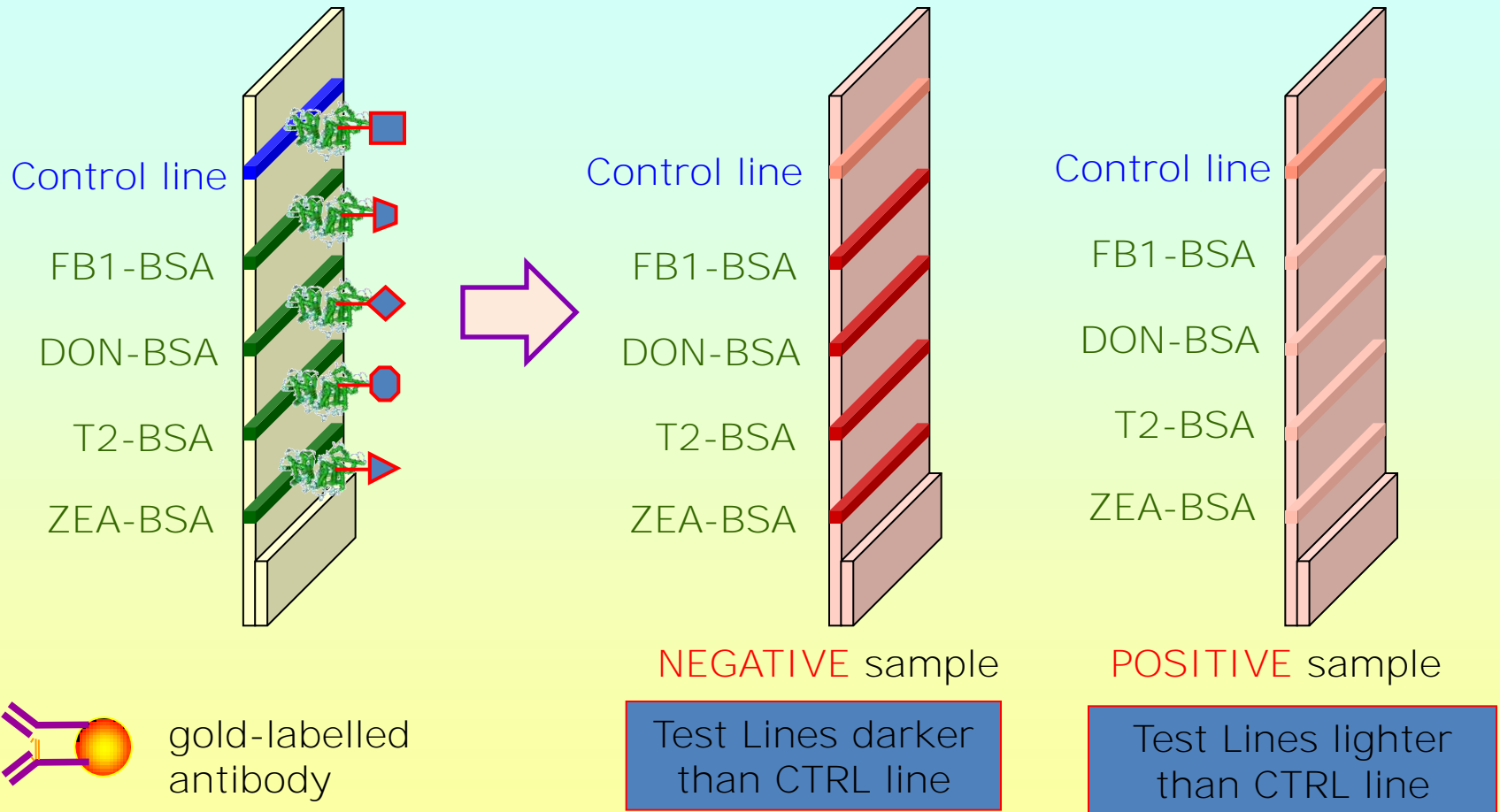


ELISA	FB₁	T-2	DON	ZEA
IC₅₀ (ng/ml)	0.5	2.5	3	0.1



Multiplex dipstick design

Indirect competitive immunoassay



Materials chosen for method development



WHEAT



OAT



WHEAT BASED EXTRUDED SNACKS

Target toxins: **DON, ZEA, T-2** and **HT-2** toxins



MAIZE



MAIZE FEED



MAIZE BASED EXTRUDED SNACKS

Target toxins: **DON, ZEA, FB₁, FB₂, T-2** and **HT-2** toxins

Required cut off: 80% of EU maximum permitted levels

Sample preparation and analysis



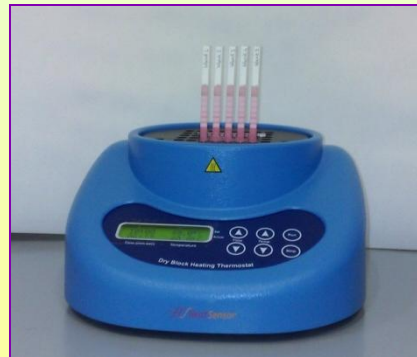
Raw cereals
Breakfast cereals
Maize feed



Methanol/water
2 min blending



Dilution and
analysis



Incubation at 40°C
Migration

Negative sample
positive ZEA
Positive ZEA/T2
Positive ZEA/T2/DON
Positive ZEA/T2/DON/FB1



METHOD PERFORMANCES

Extraction recoveries, % (RSDr %)				
	ZEA	T-2 +HT-2	DON	FB ₁ +FB ₂
WHEAT	103 (6)	73 (7)	97 (8)	-
OATS	107 (9)	73 (6)	93 (4)	-
MAIZE	105 (6)	107 (4)	105 (3)	109 (3)

Recoveries were evaluated in triplicate at cut-off levels

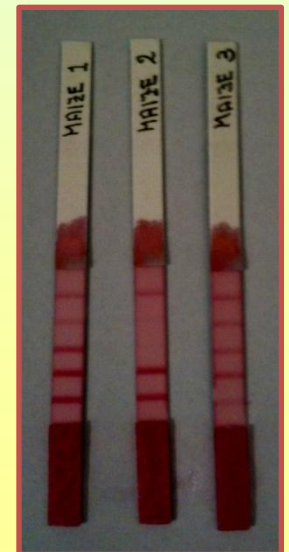
CUT OFF levels (µg/kg)				
	ZEA	T-2 +HT-2	DON	FB ₁ +FB ₂
WHEAT	80	400	1400	-
OATS	80	400	1400	-
MAIZE	280	400	1400	3200

Analysis of Naturally Contaminated Maize Samples

Sample	ZEA		T-2 +HT-2		DON		FB ₁ +FB ₂	
	dipstick	LCMSMS µg/kg	dipstick	LCMSMS µg/kg	dipstick	LCMSMS µg/kg	dipstick	LCMSMS µg/kg
1	NEG	<i>n.d.</i>	NEG	<i>n.d.</i>	NEG	<i>n.d.</i>	NEG	725
2	NEG	<i>n.d.</i>	NEG	<i>n.d.</i>	POS	24200	POS	8150
3	POS	420	LOW POS	392	LOW POS	298	NEG	725

Good agreement between dipstick and LC-MS/MS analysis.

CUT OFF levels (µg/kg)				
	ZEA	T-2 +HT-2	DON	FB ₁ +FB ₂
MAIZE	280	400	1400	3200



CONCLUSIONS (I)

LC-MS(MS) MULTI-MYCOTOXIN DETERMINATION IN FOOD

Liquid chromatography coupled to tandem MS or high resolution MS provides a reliable tool for quantitation of mycotoxins in foods at regulatory levels.

Legislation requirements are fulfilled with respect to:

- **recoveries** (EC Regulation 401/2006)
- **repeatability** and **reproducibility** (EC Regulation 401/2006)
- **LOD** enabling to detect mycotoxins at regularory levels in the concerned matrices (EC regulations *1881/2006 and 1126/2007*)
- **mass spectrometry detection** *EC performance criteria (2002/657/EC)*
(*Doc No Sanco/10684/2009*)

CONCLUSIONS (II)

LC-MS/MS DETERMINATION OF MYCOTOXIN BIOMARKERS IN HUMAN AND ANIMAL URINE

Advanced LC-MS/MS methodologies have been used for:

- ✓ **simultaneous determination of multi-mycotoxin biomarker in urine**
- ✓ **direct determination of mycotoxin metabolic profile at trace levels**

Methods based on high selective and sensitive LC-MS/MS are applicable to **monitoring programmes** providing a **complete and realistic framework of exposure levels and relevant metabolic routes.**

CONCLUSIONS (III)

MULTI-MYCOTOXIN DETERMINATION BY DIPSTICK IMMUNOASSAYS

✓ **Multiplex dipstick immunoassays** for the determination of ZEA, T-2+HT-2, DON and FB₁+FB₂ in cereals, cereal-based food and maize feed have been developed, combining the concepts of “**multiplex**” and “**rapid**” detection.

✓ The resulting immunoassay protocol is: **rapid, inexpensive,** and **easy-to-use.**

✓ **Robustness and reliability of dipstick based methods should be demonstrated through interlaboratory studies.**

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