

# DESI-MS

## in food contaminants control

Michel Nielen, Hans Mol



# Outline

Introduction

Principle

Experimental parameters

Instrumentation

Food contaminant applications

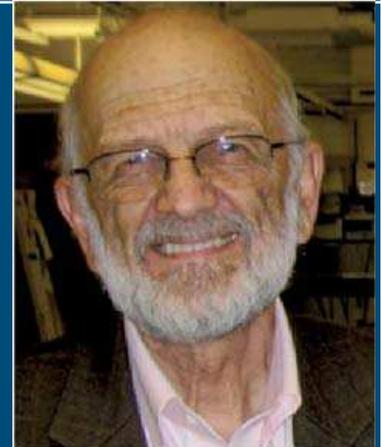
Summary/outlook

# DESI-MS

**D**esorption **e**lectrospray **i**onization

First ambient MS technique invented by Graham Cooks from Purdue University, USA

**M**ass **s**pectrometry

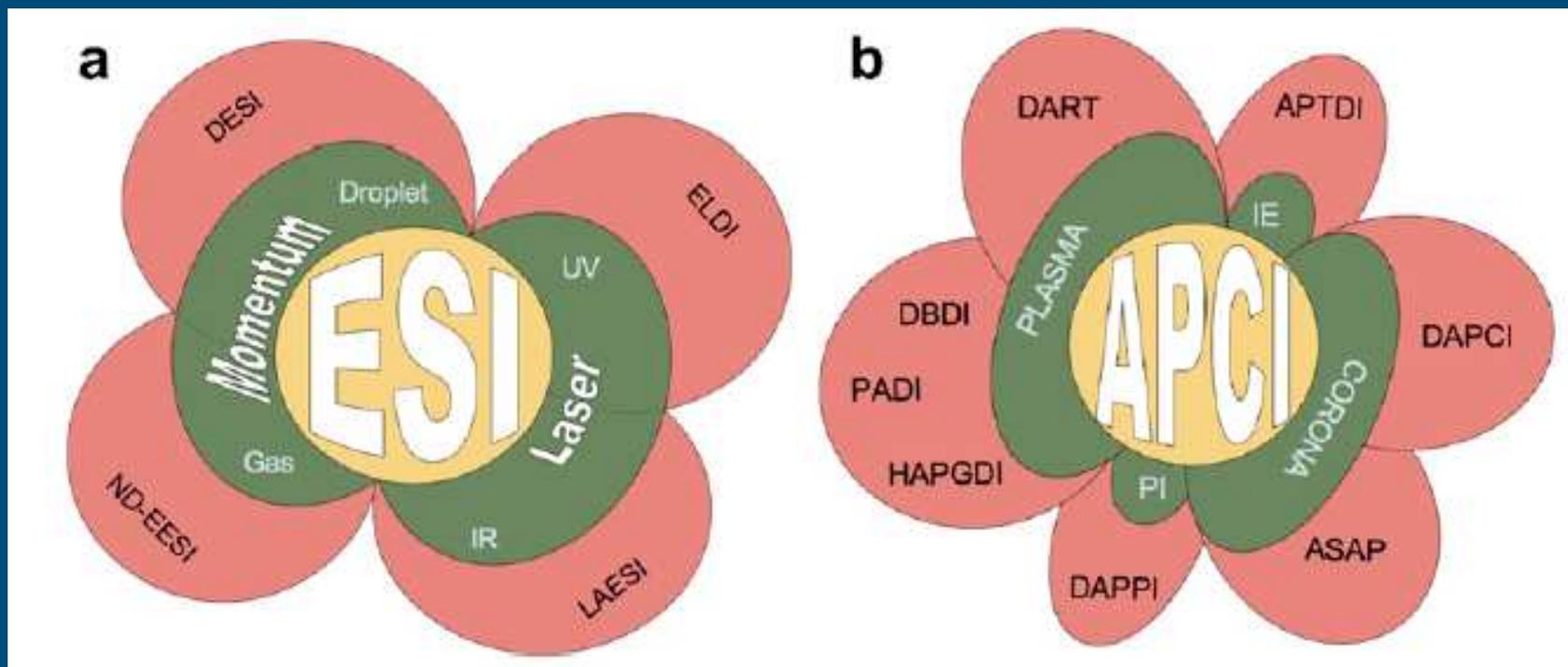


## Mass Spectrometry Sampling Under Ambient Conditions with Desorption Electrospray Ionization

Zoltán Takáts, Justin M. Wiseman,  
Bogdan Gologan, R. Graham Cooks\*

Science 306, 471 (2004)

# Ambient MS techniques



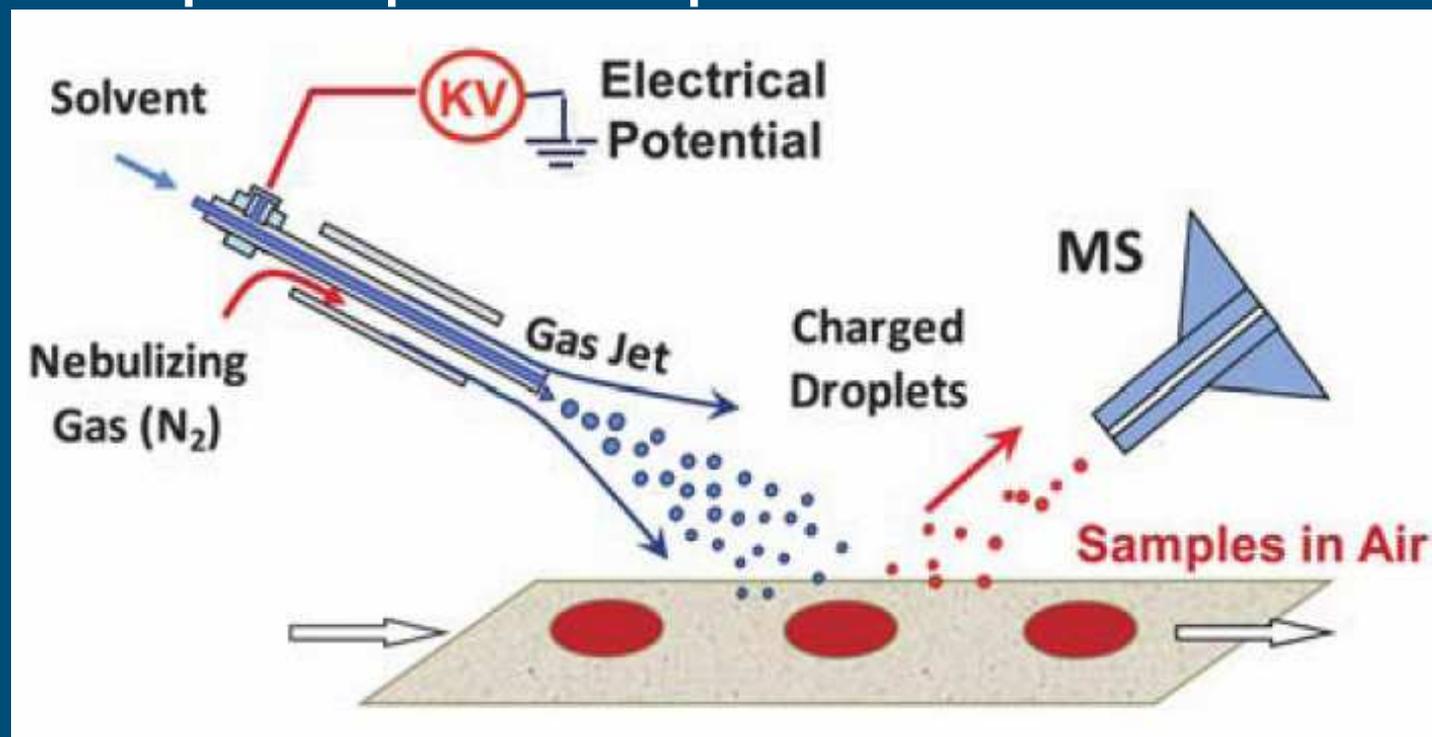
Venter, Nefliu, Cooks, Trends in Anal. Chem. 27 (2008) 284-290

# Ambient MS

**Table 1** Chronology and perceived impact of developments in ambient ionization MS **Weston, Analyst, 2010, 135, 661-668**

Method name	Acronym or abbreviation	Year of introduction	Research papers (papers/year) <sup>a</sup>
Desorption electrospray ionization	DESI <sup>1</sup>	2004	188 (31)
Surface sampling probe	SSP <sup>20</sup>	2004	5 (1)
Direct analysis in real time	DART <sup>2</sup>	2005	65 (16)
Atmospheric solids analysis probe	ASAP <sup>21</sup>	2005	7 (1.4)
Electrospray laser desorption ionization	ELDI <sup>22</sup>	2005	6 (1.5)
Fused droplet electrospray ionization	FD-ESI <sup>23</sup>	2005	2 (0.5)
Direct atmospheric pressure chemical ionization	DAPCI <sup>24</sup>	2005	89 (22)
Matrix-assisted laser desorption electrospray ionization	MALDESP <sup>25</sup>	2006	8 (2.7)
Jet desorption electrospray ionization	JeDI <sup>26</sup>	2006	1 (0.3)
Extractive electrospray ionization	EESI <sup>27</sup>	2006	22 (7)
Desorption sonic spray ionization	DeSSI <sup>28</sup>	2006	2 (0.7)
Atmospheric pressure thermal desorption ionization	APTDI <sup>29</sup>	2006	3 (1)
Helium atmospheric pressure glow discharge ionization	HAPGDI <sup>30</sup>	2006	1 (0.5)
Plasma-assisted desorption ionization	PADI <sup>31</sup>	2007	1 (0.5)
Dielectric barrier desorption ionization	DBDI <sup>32</sup>	2007	2 (1)
Neutral desorption extractive electrospray ionization	ND-EESP <sup>33</sup>	2007	5 (2.5)
Laser diode thermal desorption	LDTD <sup>34</sup>	2007	1 (0.5)
Laser ablation electrospray ionization	LAESI <sup>35</sup>	2007	1 (0.5)
Desorption atmospheric pressure photo-ionization	DAPPI <sup>36</sup>	2007	2 (1)
Infra red laser ablation electrospray ionization	IR-LAESI <sup>37</sup>	2008	7 (3.5)
Flowing atmospheric-pressure afterglow	FAPA <sup>38</sup>	2008	2 (2)
Easy ambient sonic spray ionization	EASI <sup>39</sup>	2008	5 (5)
Remote analyte sampling transport and ionization relay	RASTIR <sup>40</sup>	2008	1 (1)
Laser ablation flowing atmospheric-pressure afterglow	LA-FAPA <sup>41</sup>	2008	5 (5)
Low temperature plasma	LTP <sup>42</sup>	2008	7 (7)
Desorption electrospray metastable-induced ionization	DEMI <sup>43</sup>	2009	1 (-)
Liquid micro-junction surface sampling probe/electrospray ionization	LMJ-SSP/ESI <sup>44</sup>	2009	3 (-)
Surface activated chemical ionization	SACI <sup>45</sup>	2009	1 (-)
Single particle aerosol mass spectrometry	SPAMS <sup>46</sup>	2009	1 (-)

# DESI: principle and processes



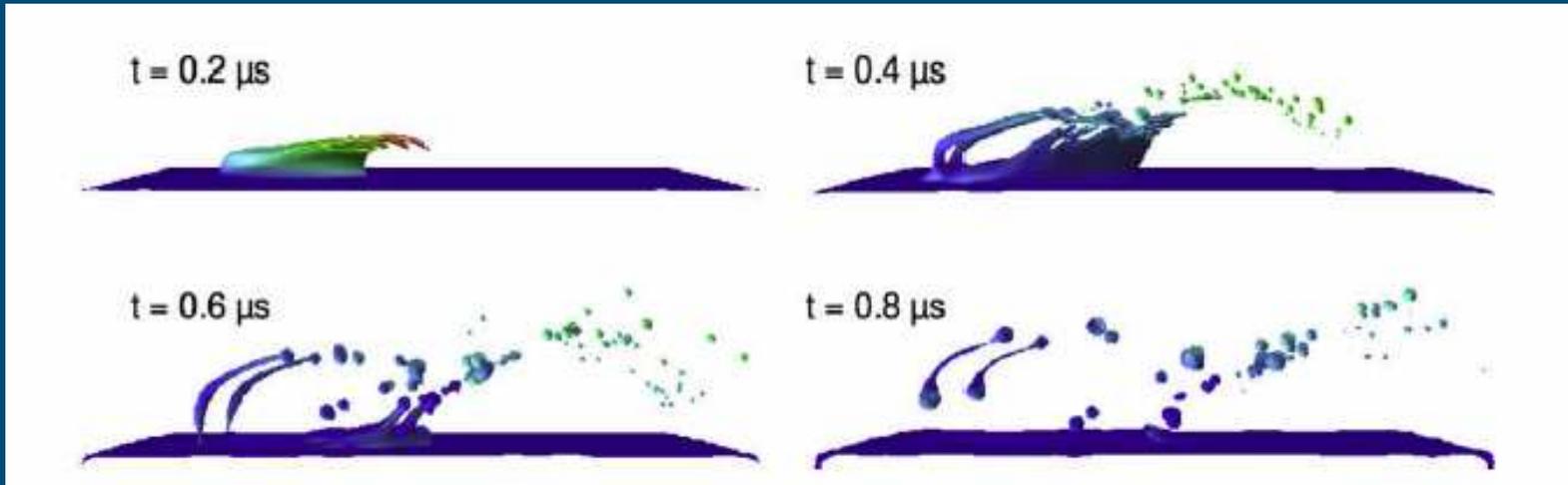
## From surface deposit to ion:

1. Formation of charged primary droplets by electrospray source
2. Flight of droplets to surface, wetting of surface, dissolution
3. Collision of charged droplets with surface => smaller droplets
4. Takeoff of secondary droplets, ablated material, free ions
5. Transport through atm. pressure interface
6. Shrinking of droplets, electrostatic fission => free ions

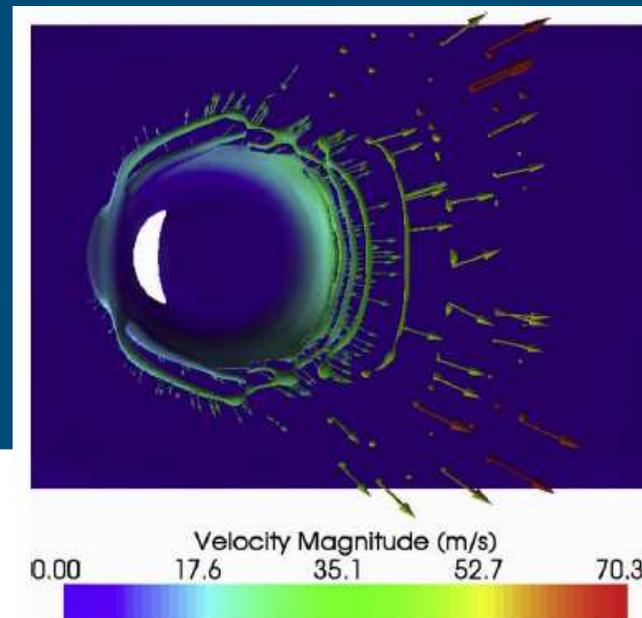
Ifa et al, Analyst, 2010, 135, 669–681



# Formation of secondary droplets

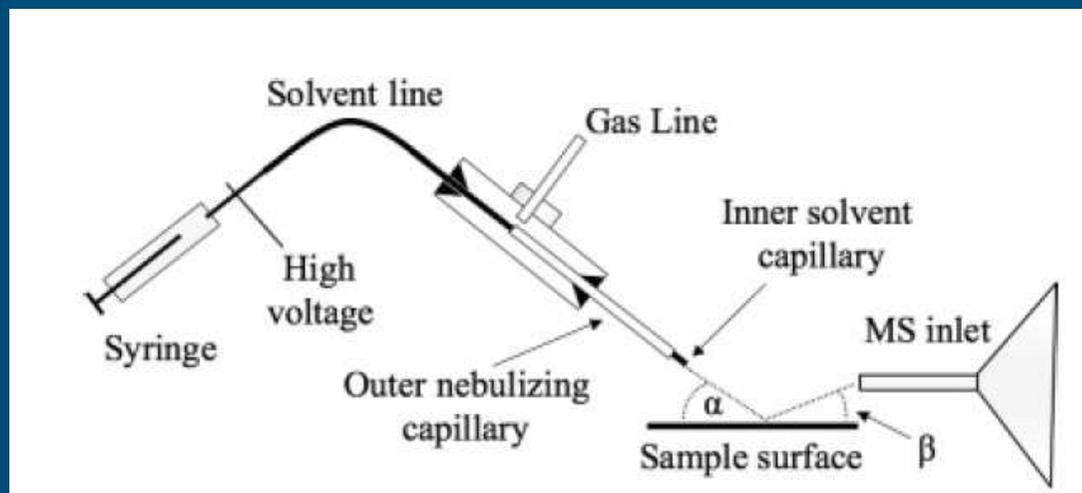


Costa, Cooks, Chemical Physics Letters 464 (2008) 1–8



# Experimental parameters

- Solvent
- Solvent flow
- Gas flow
- Spray angle
- Capillary voltage
- Distance to sample surface
- Angle ion collection MS
- Sample surface
- Distance to MS inlet
- MS parameters (incl. source temp)



# Solvent and spray

Dissolution of analyte in microdroplets / solvent film

Ionization; similar to LC-ESI-MS solvents

⇒ MeOH or ACN with water/acid

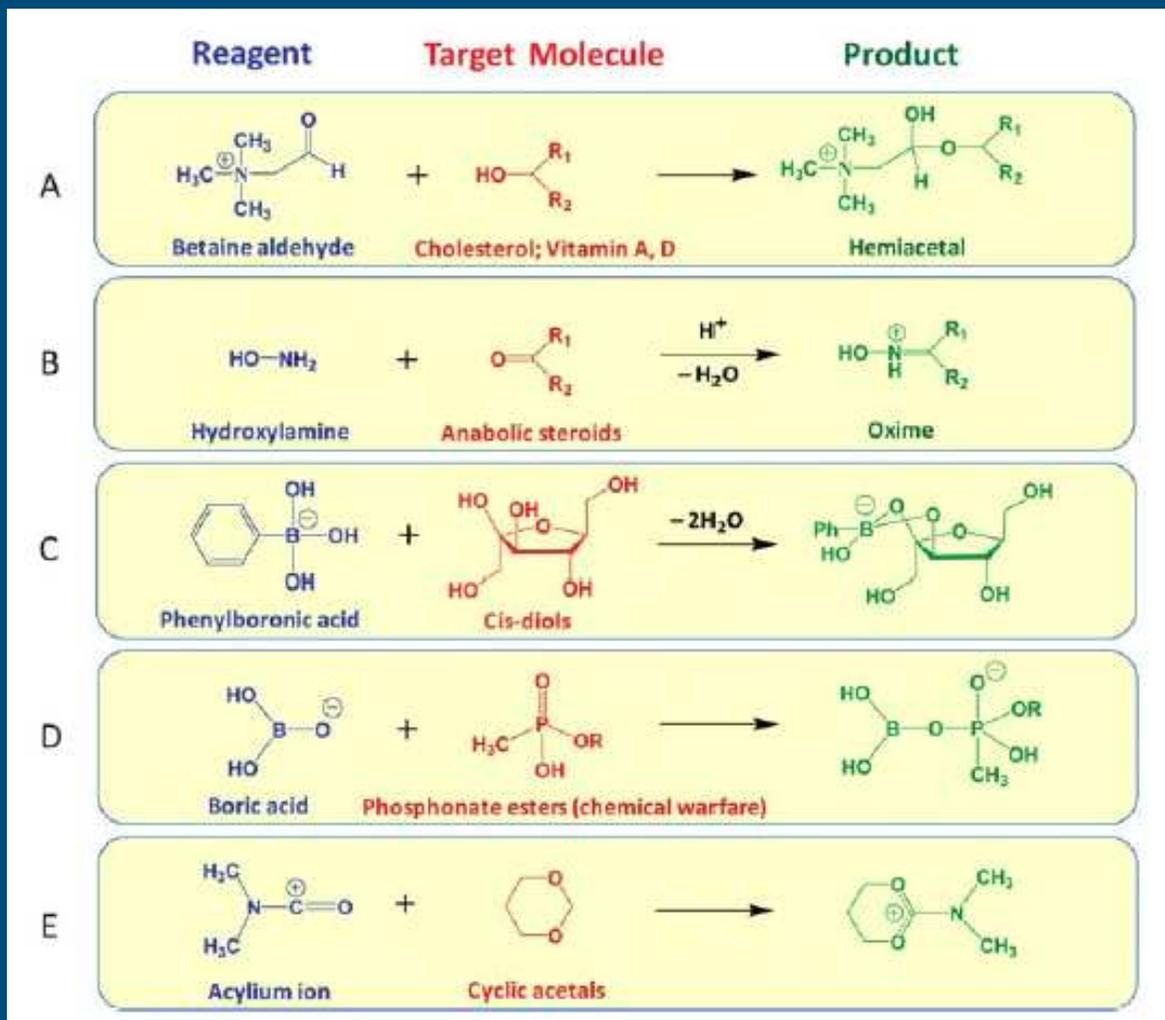
Option to add reagents (reactive DESI)

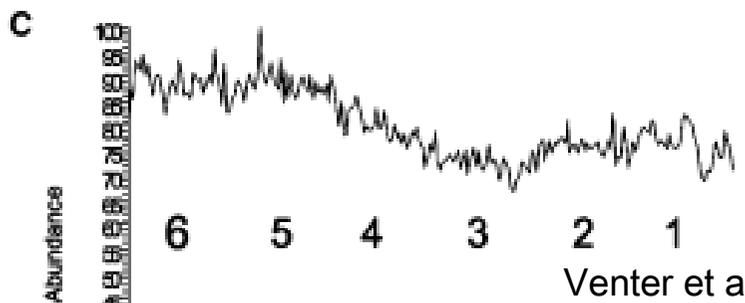
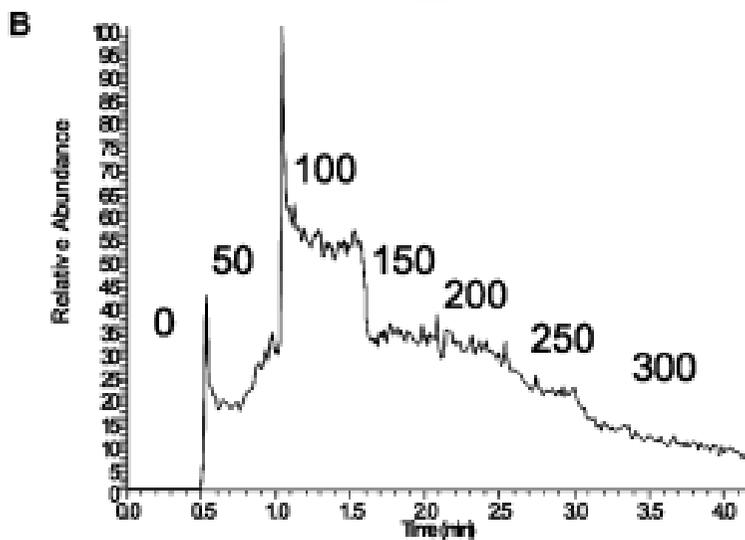
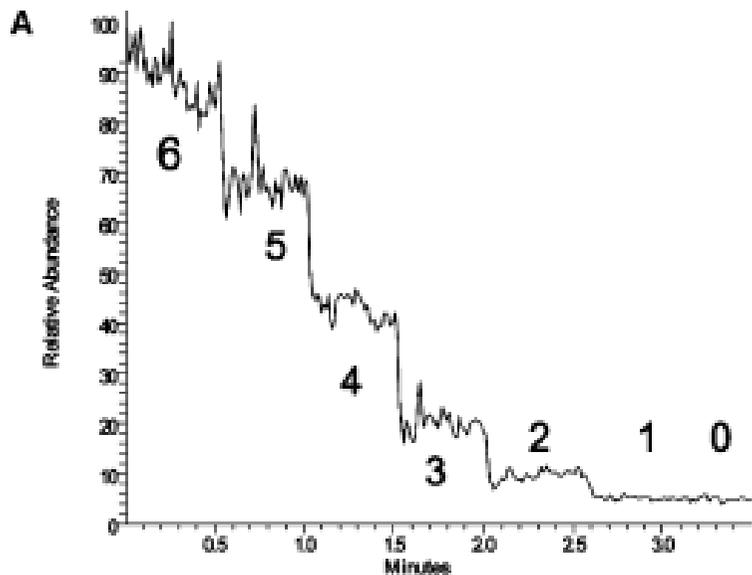
Flow rate: 1-5  $\mu\text{l}/\text{min}$

Nebulizer gas: 8-40 L nitrogen/min

Spray droplets:  $\leq 10 \mu\text{m}$  (2-4),  $v \geq 100 \text{ m/s}$  (120)

# Reactive DESI





Spray voltage (kV)

Higher  $V \Rightarrow$  more charges on droplet surface

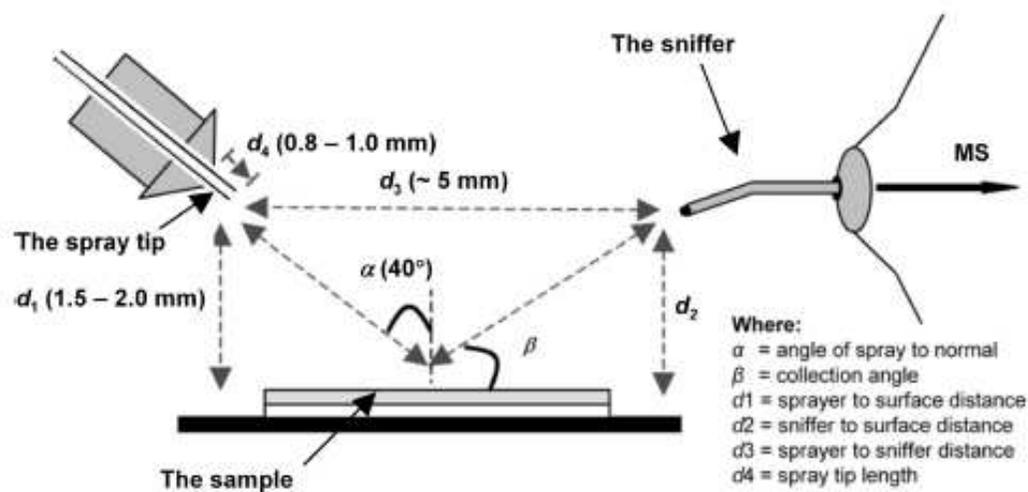
Levels off around 5 kV ( $\Rightarrow$  default)

Nebulizer gas pressure (psi)

Solvent flow rate  $\mu\text{l}/\text{min}$

# Geometry

## Standard configuration DESI sources



Green et al, Anal. Chem. 2009, 81, 2286-2293

$\alpha = 25-80^\circ$

$\beta = \sim 5-10^\circ$

Spray tip to surface = 1-10 mm

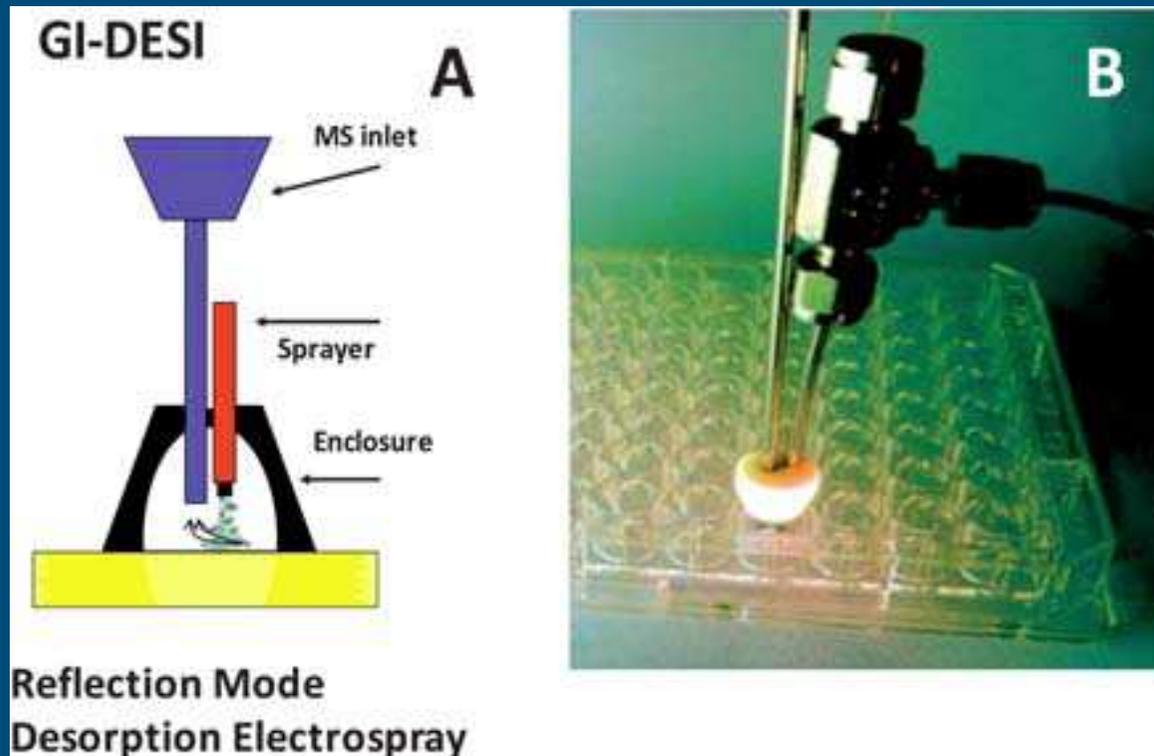
Surface to MS inlet = 1-5 mm

## Alternative configurations

Configuration	Incident/Collection angle	Mean Rhodamine intensity*
	90°/90°	1546±630
	90°/10°	739±250
	50°/10°	1375±510
	45°/45°	2974±1040
	50°/10°(Open)	1490±525

Venter et al, Anal. Chem. 2007, 79, 6398

# Geometry independent configurations



# Sample surface (substrate)

Wettability: solvent formation/dissolution

Electroconductivity: more charge maintained by substrate, more charge in sec. droplets

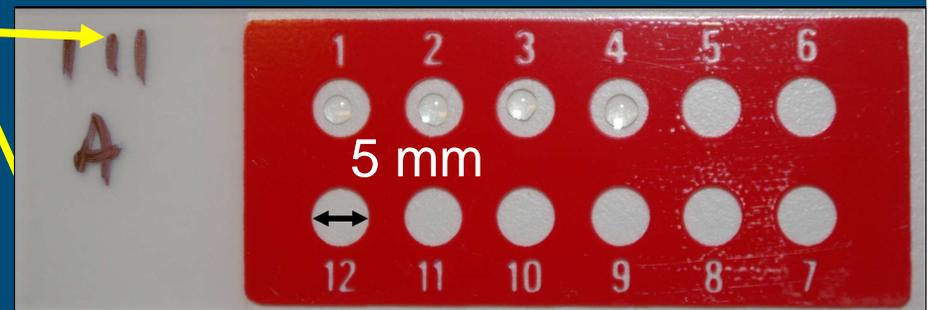
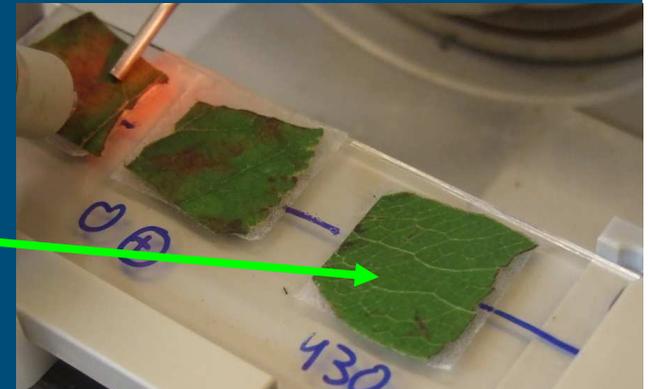
## Surfaces:

Sample itself

Substrates for liquids/extracts:

- Glass
- PTFE (teflon)
- PMMA (polymethylmethacrylate)
- Filter paper
- TLC plates

Pear leaf

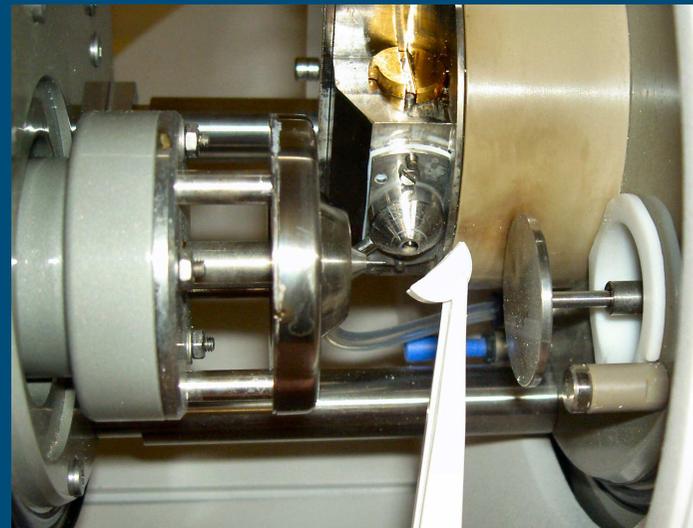


# DESI, instrumentation

In principle.....you can misuse a Z-source....



Paracetamol



# DESI, instrumentation

## Ion source assembly

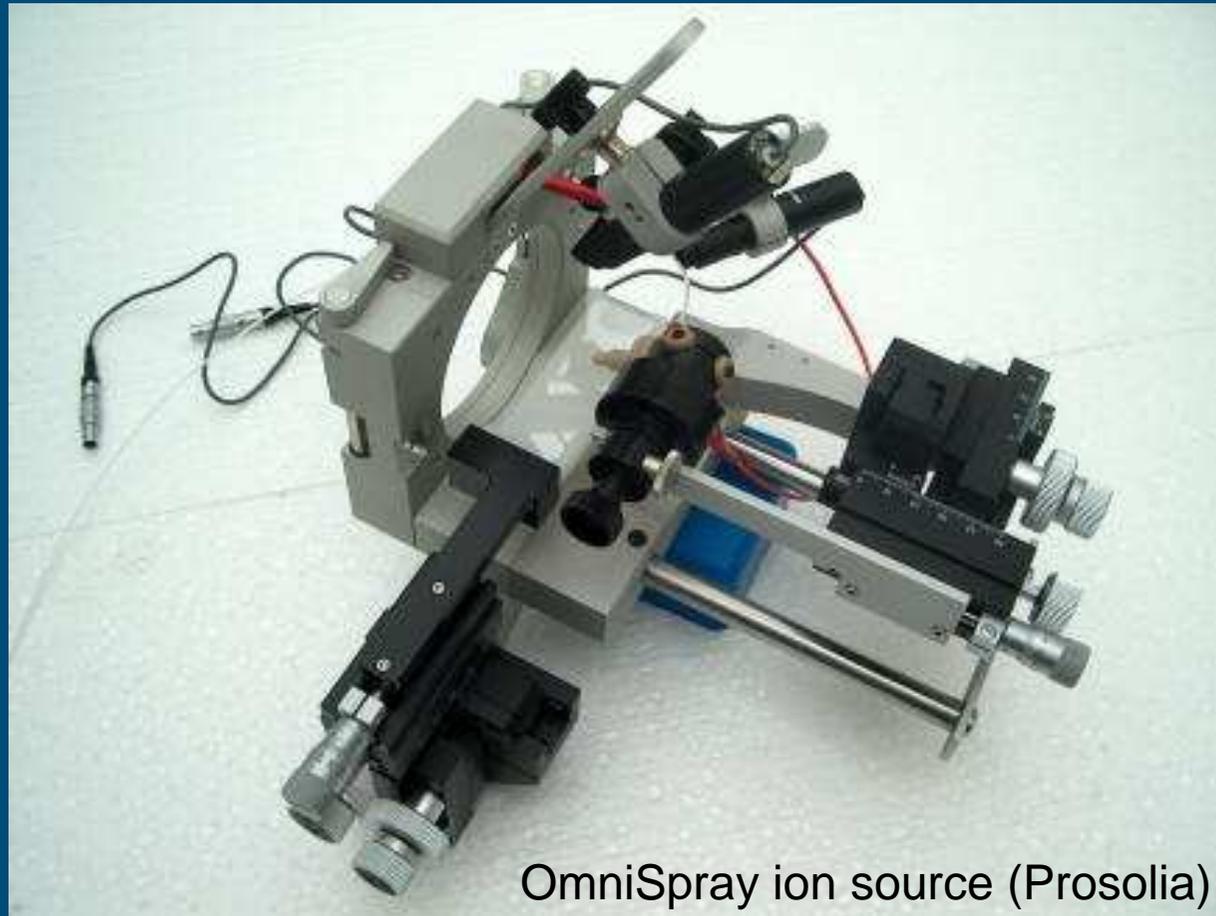
## Mass spectrometer

OmniSpray ion source



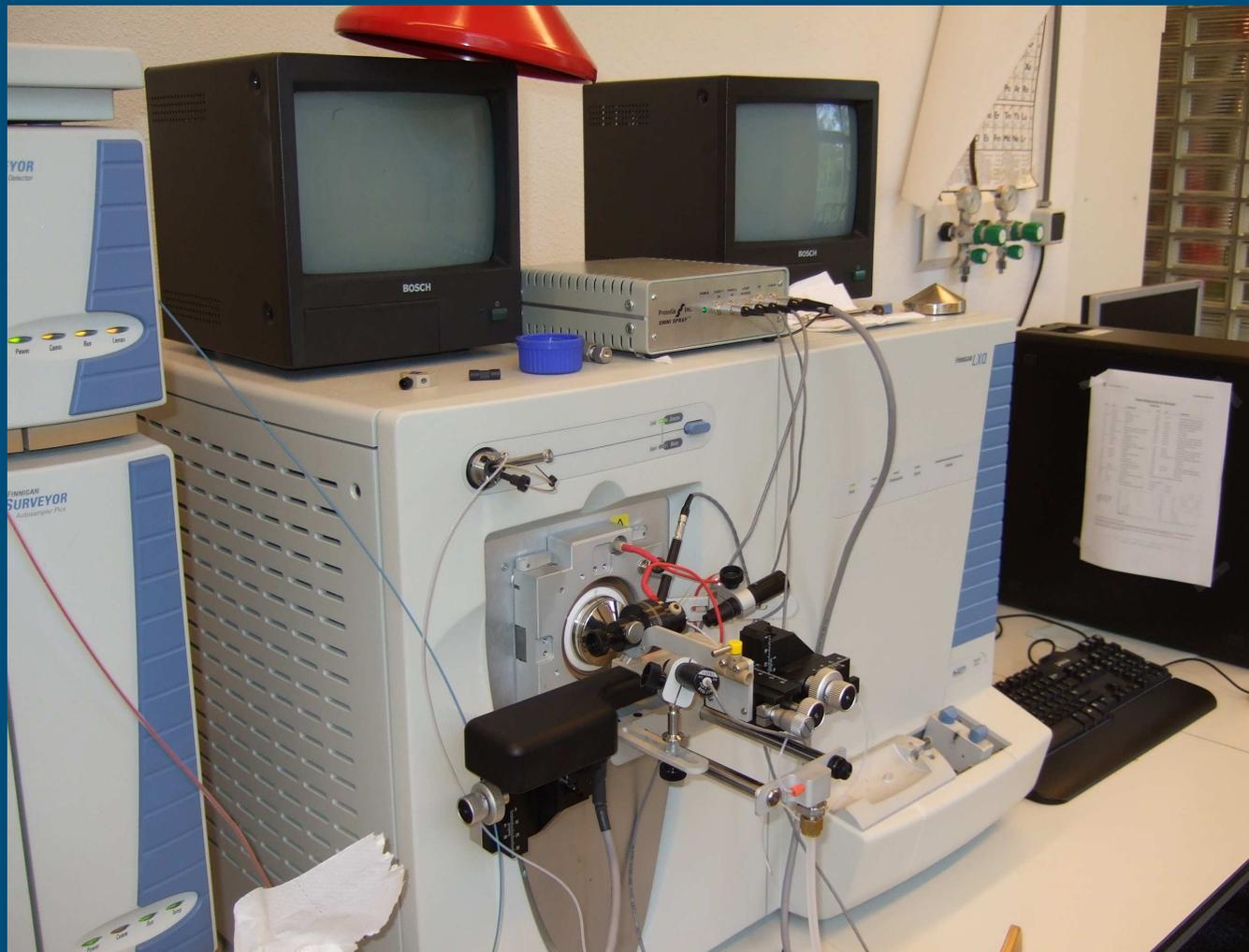
# DESI, instrumentation

## Top view DESI source assembly

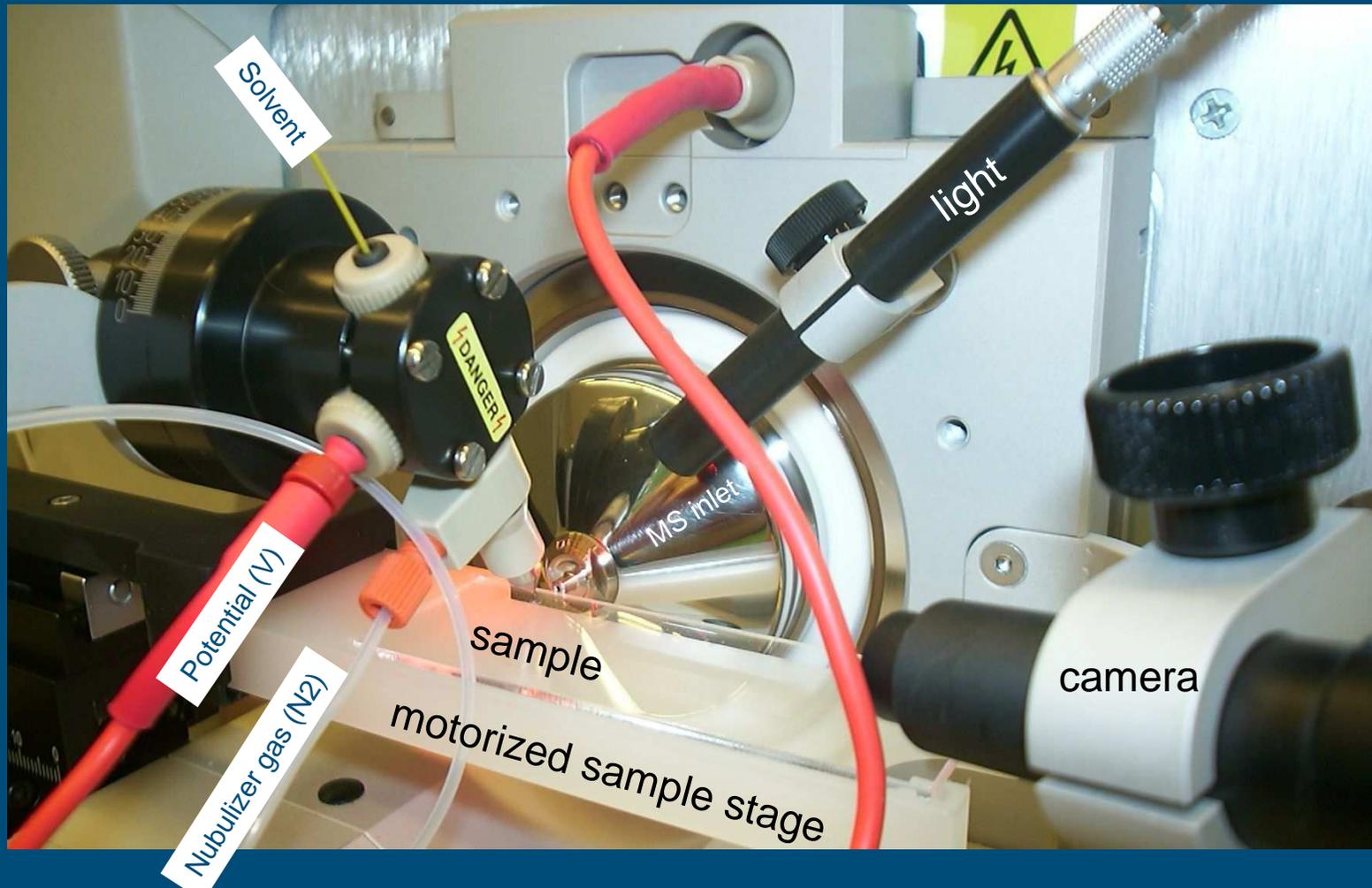


OmniSpray ion source (Prosolia)

# DESI-MS/MS (LXQ, linear ion trap)

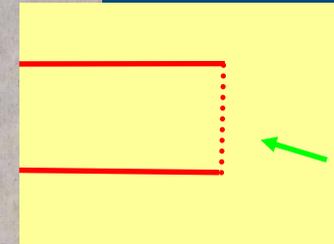


# DESI, instrumentation

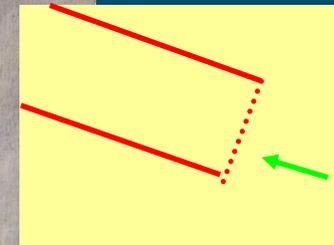


# DESI, MS inlet (sniffers)

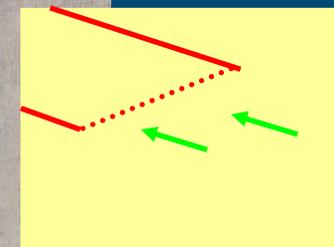
Standard LXQ MS inlet capillary



Extended, bended

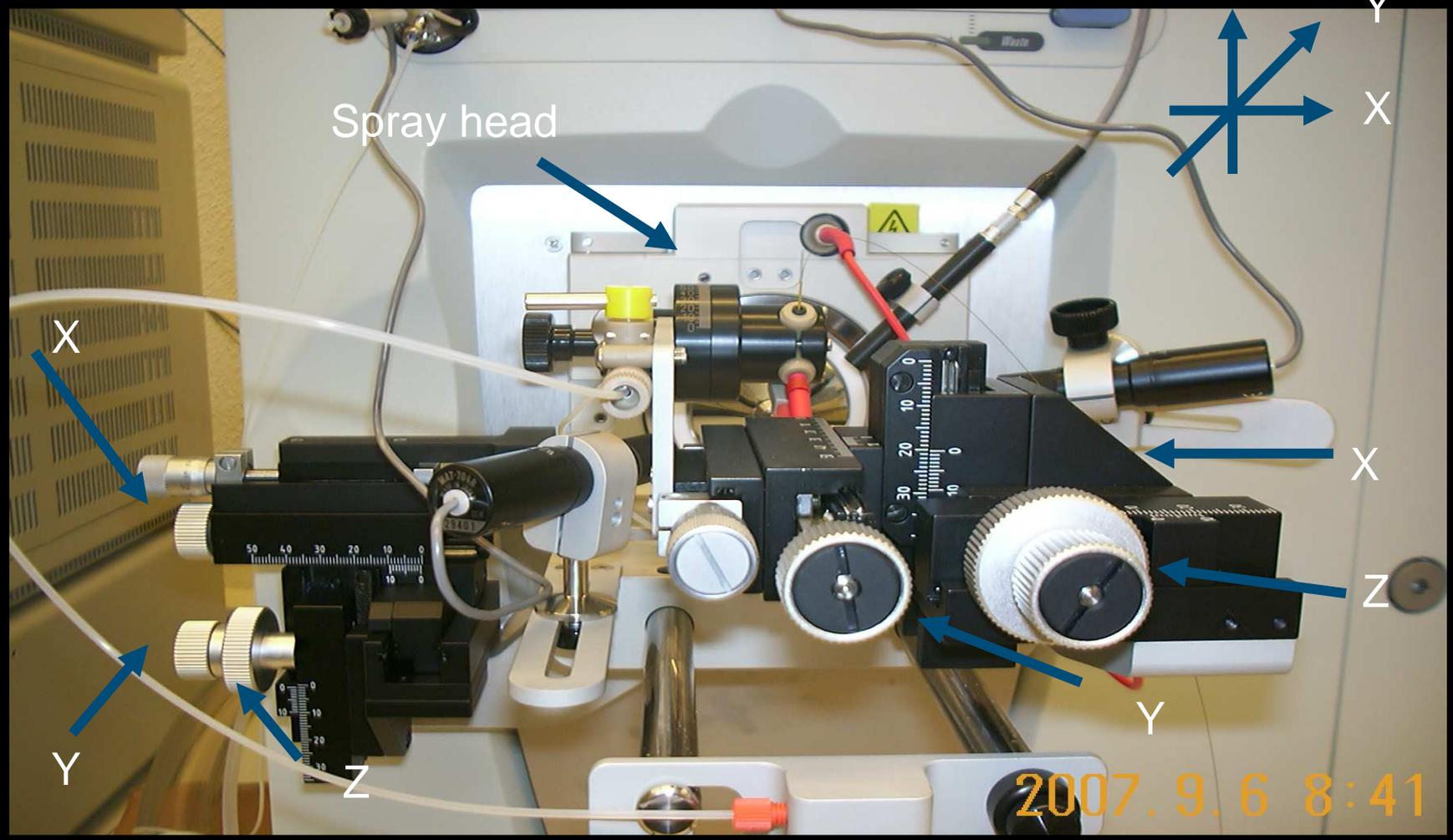


Extended, bended, enlarged opening





# Front view DESI



# DESI applications

DESI and related ESI approaches by far most utilized ambient MS technique\*

Rapid analysis pharmaceutical product

Explosives

Biological imaging

Forensic imaging of inks

Illegal and counterfeit drugs

Drug metabolites in biological matrices

Chemical warfare agents

Textiles

Peptides and proteins

Proteomics

Metabolomics

Plants / natural products

**Food contaminants**

.....

# DESI in food contaminants control

## Consideration MRL verification

- ⇒ MRLs expressed in mg/kg whole product
- ⇒ requirements minimum sample size

## Test MRL compliance:

Homogenisation (+extraction) => deposit on spots/slides  
(semi) Quantitative analysis

**In laboratory**  
DESI for fast  
high throughput  
analysis of extracts

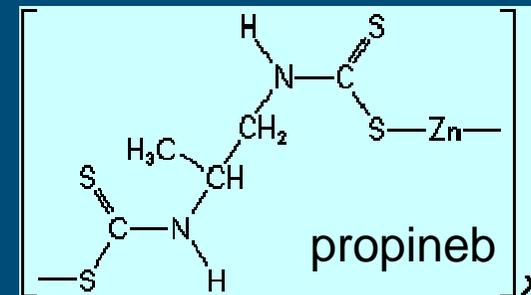
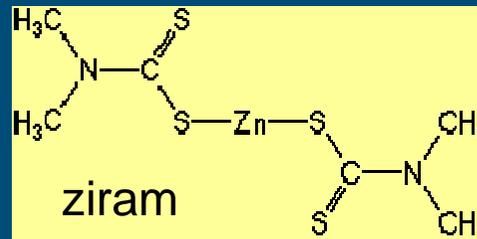
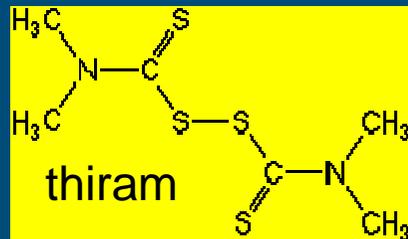
## Test presence (screening, yes/no)

Direct detection from sample surface  
or cross-sections of tissues  
No sample preparation  
Qualitative

**In laboratory**  
Fast high throughput analysis of sample  
Distribution on surface/cross sections  
**In-field**  
DESI as on-site detection tool  
Food forensics  
crops, swops, clothing, containers, syringes

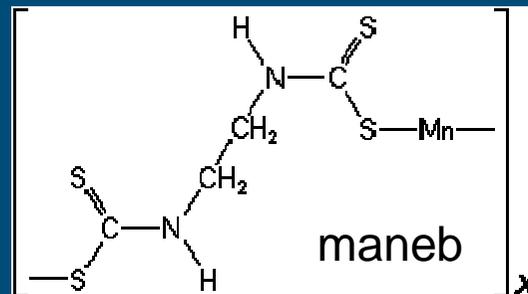
# Feasibility Dithiocarbamates by DESI

Fungicides, widely used, high application rates  
Non-systemic (residues on surface only)



DMDC = bis(dimethyldithiocarbamate)

PBDC =  
propylenebis(dithiocarbamate)



EBDC = ethylenebis(dithiocarbamate)



# Dithiocarbamates

Solubility issues

Stability issues

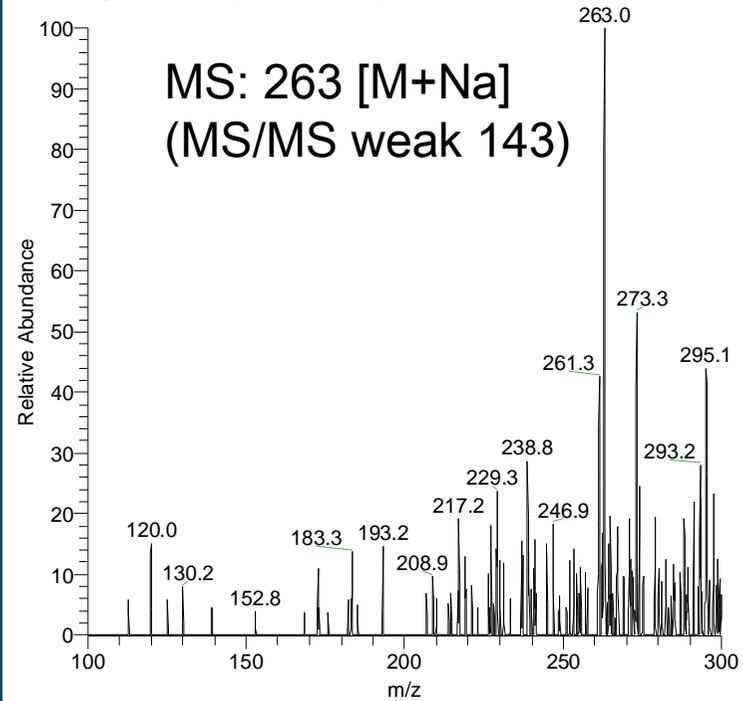
Current method: hydrolysis to CS<sub>2</sub>

Legislation (396/2005): CS<sub>2</sub> (sum) but specific MRL for 3 DTCs

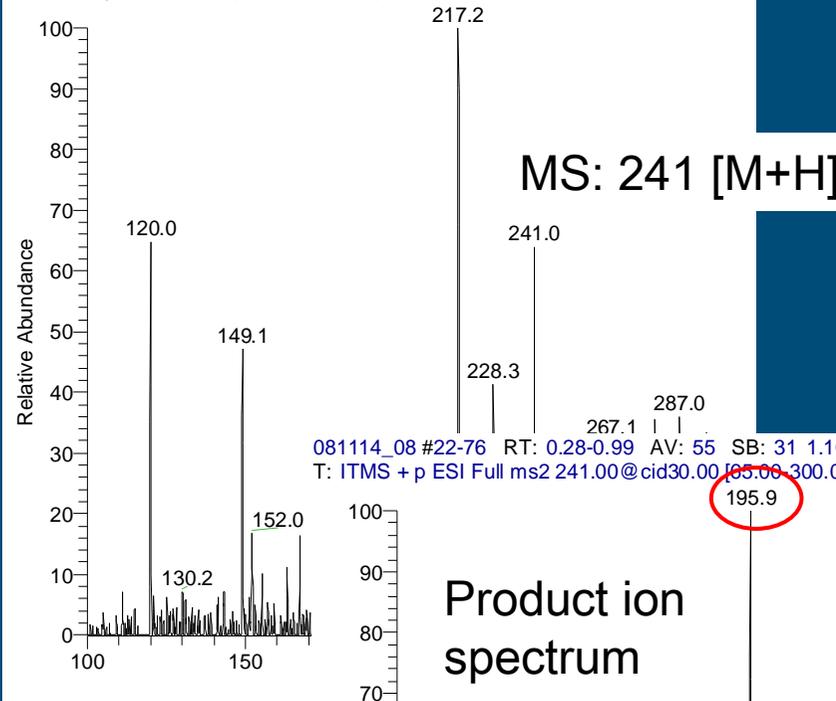
Groups and examples of individual products to which the MRLs apply (a)	Dithiocarbamates (dithiocarbamates expressed as CS <sub>2</sub> , including mancozeb, metiram, propineb, thiram and ziram)			
	Propineb (expressed as propilendiamine)	Thiram (expressed as thiram)	Ziram	
<b>Apples</b>	<b>5</b>	<b>0,3</b>	<b>5</b>	<b>0,1*</b>
<b>Pears</b>	<b>5</b>	<b>0,3</b>	<b>5</b>	<b>1</b>
<b>Strawberries</b>	<b>10 (ft)</b>	<b>0,05*</b>	<b>10</b>	<b>0,1*</b>
<b>Lettuce</b>	<b>5</b>	<b>0,05*</b>	<b>2</b>	<b>0,1*</b>

# MS/MS optimization (thiram)

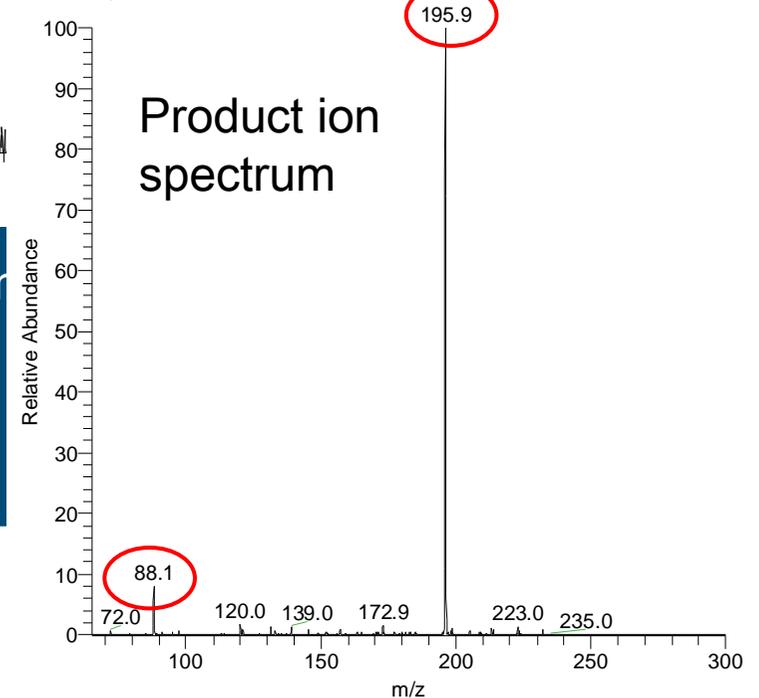
081114\_01 #56 RT: 0.54 AV: 1 NL: 3.20E1  
T: ITMS + p ESI Full ms [100.00-500.00]



081114\_02 #8-22 RT: 0.07-0.21 AV: 15 NL: 6.40  
T: ITMS + p ESI Full ms [100.00-500.00]



081114\_08 #22-76 RT: 0.28-0.99 AV: 55 SB: 31 1.10-1.50 NL: 8.01  
T: ITMS + p ESI Full ms2 241.00@cid30.00 [85.00-300.00]



MeOH/water 1:1 , 0.1% FA

.....+ 10 m

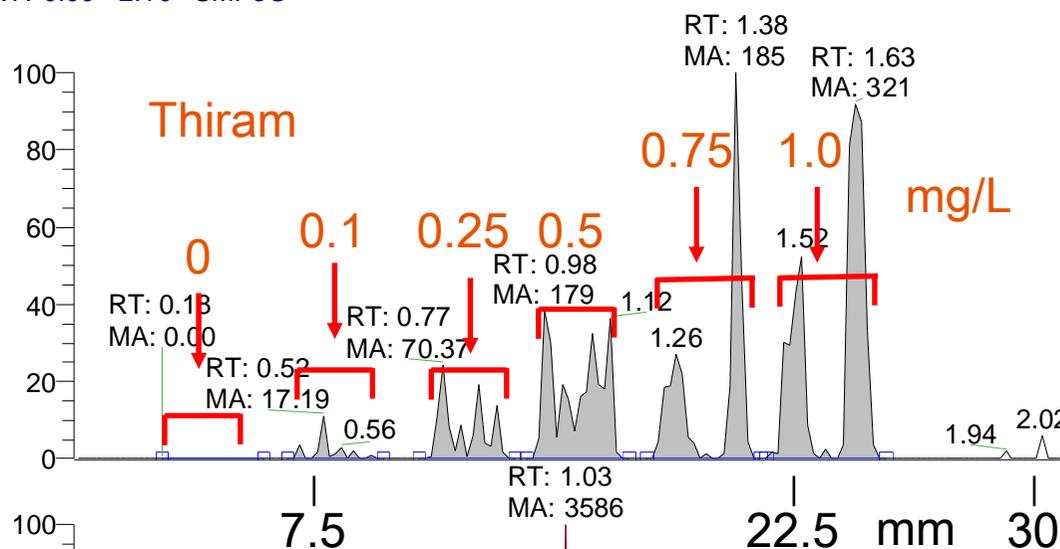
# DESI parameters

Nitrogen sheath gas	120 psi (9 bar)
Incident angle	55°
Spray tip-to-surface distance	5 mm
Surface to MS-inlet distance	1 mm
Scattering angle	ca. 10° to surface
DESI spray	MeOH/water 1:1 + 10 mM NH <sub>4</sub> For. + 0.1% FA
Flow	5 µl/min
Scanning of the surface	250 µm/sec
Heated capillary MS (LXQ)	50°C
Substrate	PTFE printed on glass

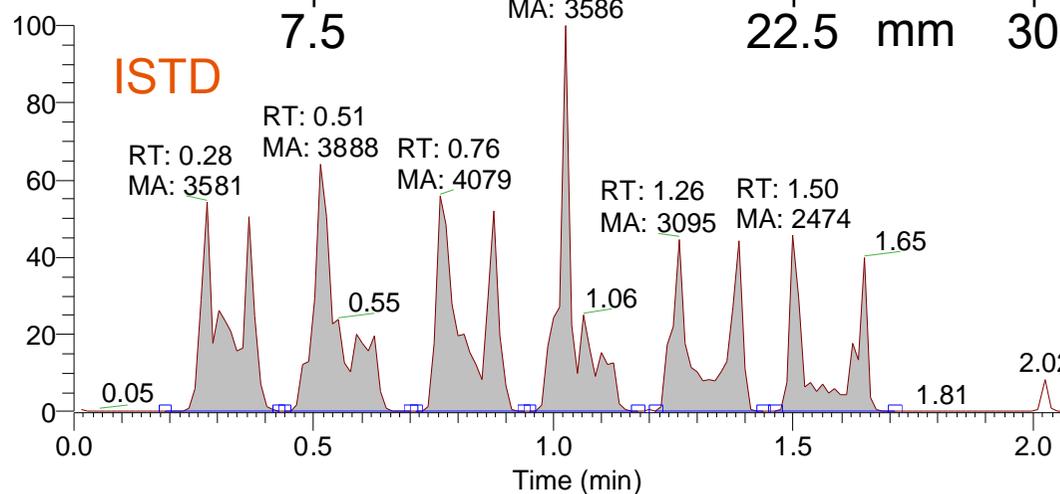
# Thiram std solutions (teflon spots on glass)



RT: 0.00 - 2.10 SM: 3G

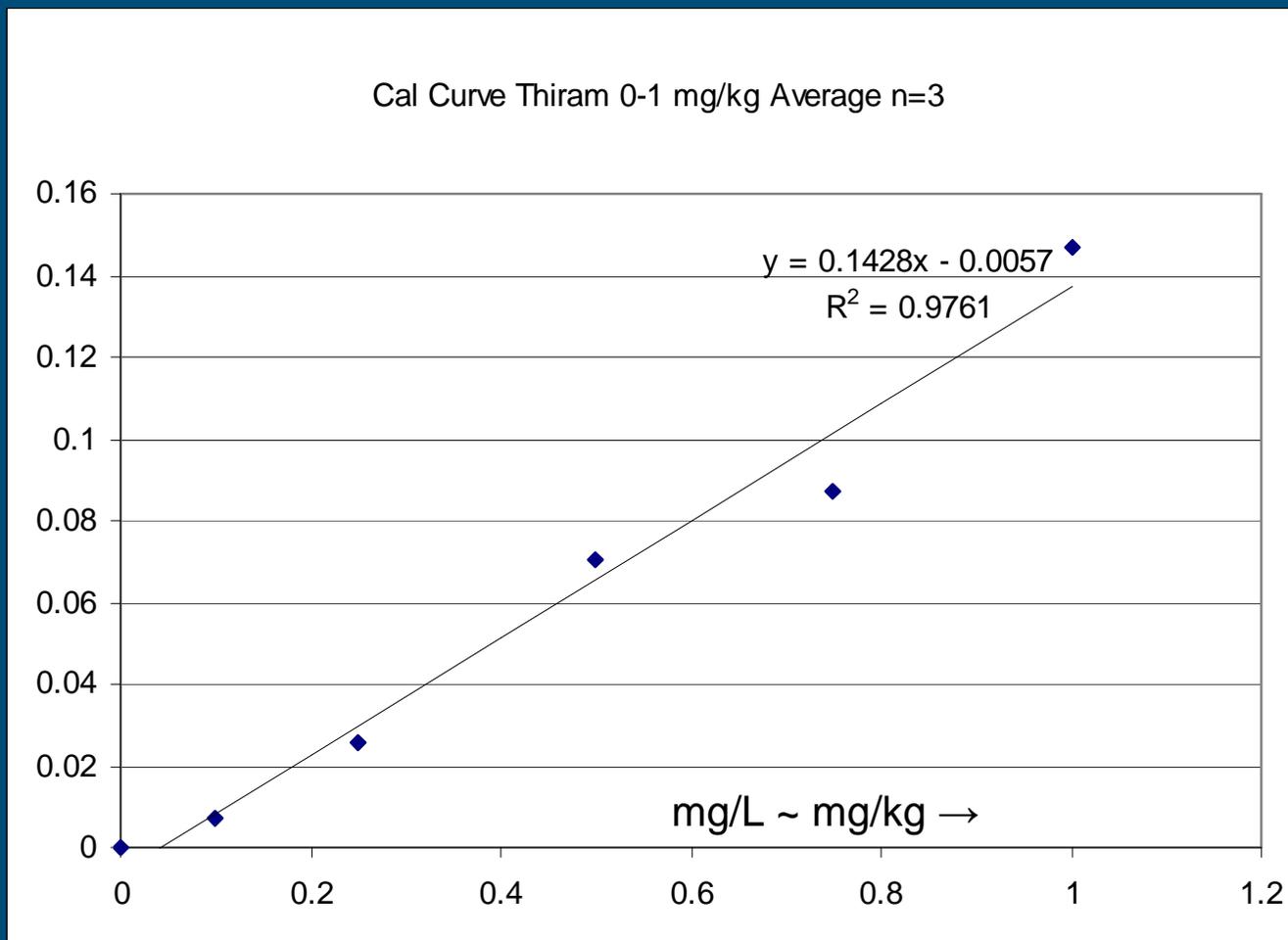


NL: 9.07E1  
 m/z=  
 195.50000-196.50000 F:  
 ITMS + c NSI Full ms2  
 241.00@cid25.00  
 [80.00-250.00] MS  
 100422\_31

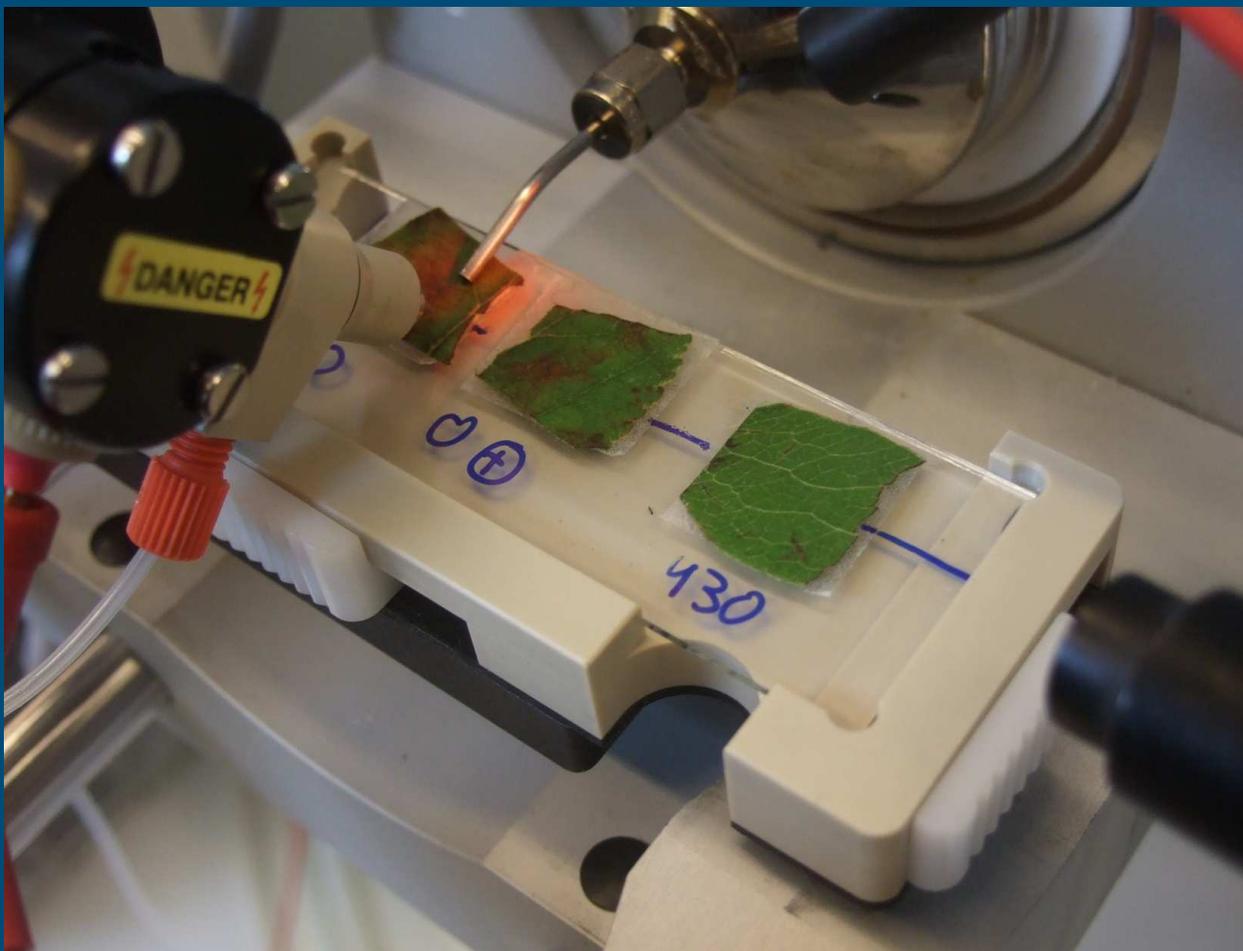


NL: 1.62E3  
 m/z=  
 371.50000-372.50000 F:  
 ITMS + c NSI Full ms2  
 404.00@cid25.00  
 [110.00-450.00] MS  
 100422\_31

# Quantitative analysis?



# Direct analysis from sample surface



# Alternative to direct analysis

Homogenization (plant juices); simple extraction procedure followed by rapid DESI detection

Options extraction:

Systemic: QuEChERS or other

Non-systemic: solvent rinse of intact product

# QuEChERS: 1. Initial Extraction Step



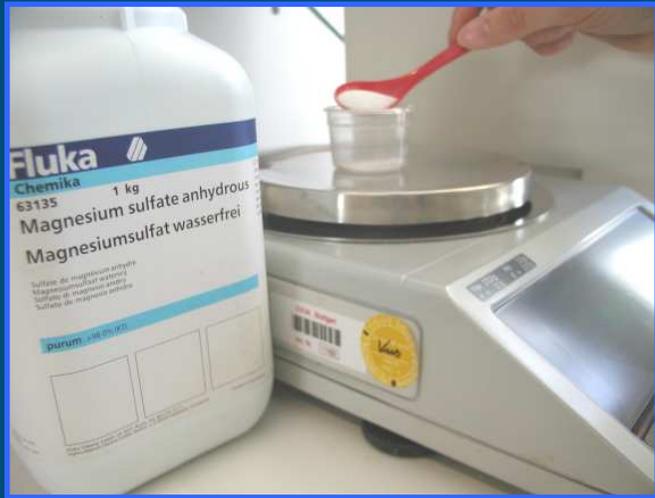
**Weigh 10 g Sample**  
**Add 10 mL MeCN**



**Shake Intensively**  
**for 1 min**



## 2. Extraction/Partitioning Step



(Pre-)Weigh  
4 g  $\text{MgSO}_4$  + 1 g NaCl

Add to the Tube



Shake Intensively  
for 1 min



### 3. Addition of ISTD and Centrifugation



Add ISTD  
Shake for 30 s



Centrifuge (ca. 5 min)  
Separated Raw Extract



## 4. Dispersive SPE Step



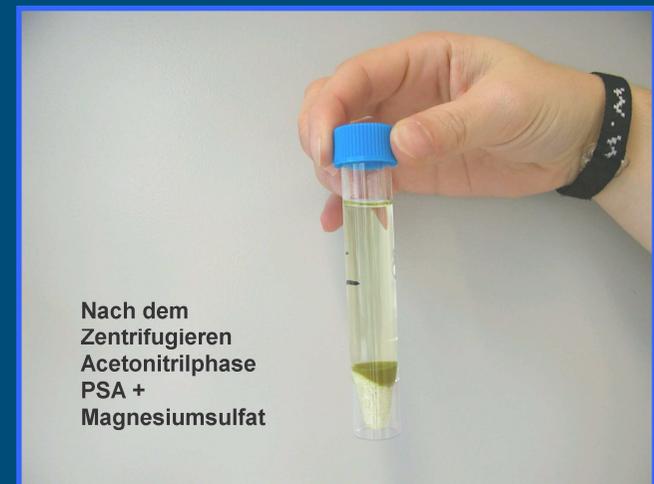
(Pre-) Weigh  
 $\text{MgSO}_4$  and PSA

Add Extract to Tube  
and Shake ca. 30 s



Centrifuge (ca. 2 min)

Cleaned up Extract

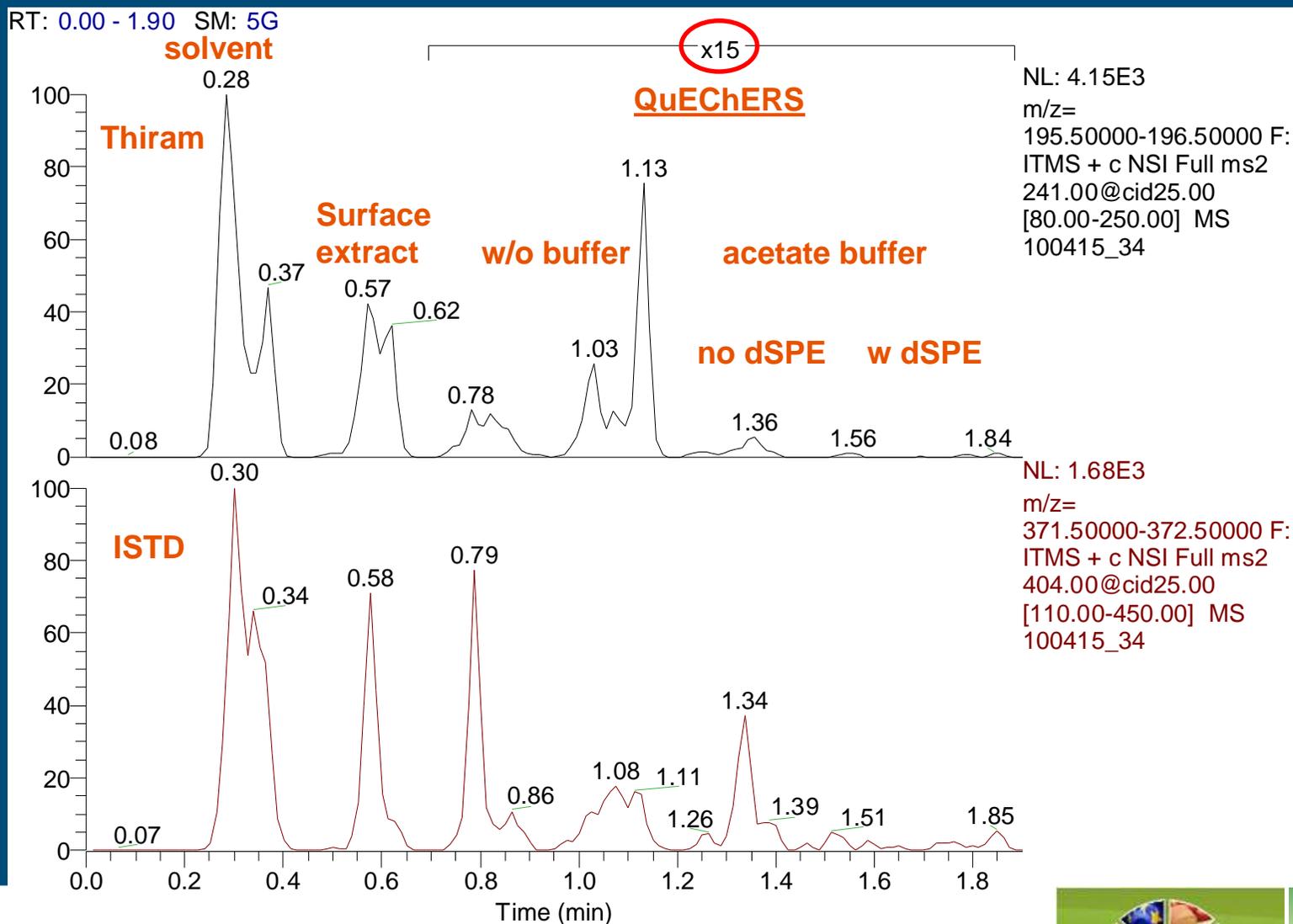


Nach dem  
Zentrifugieren  
Acetonitrilphase  
PSA +  
Magnesiumsulfat

# Surface extraction

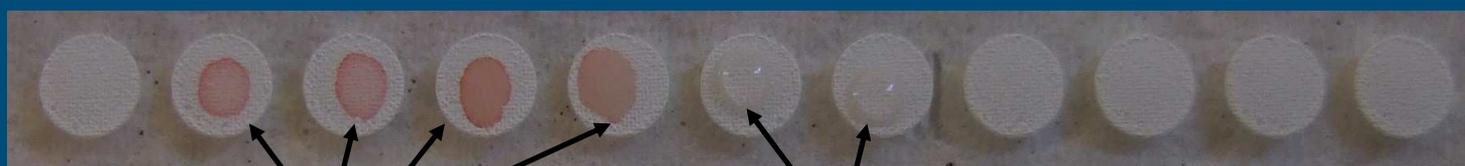


# Comparison of extraction methods (pear)



# Spike experiments

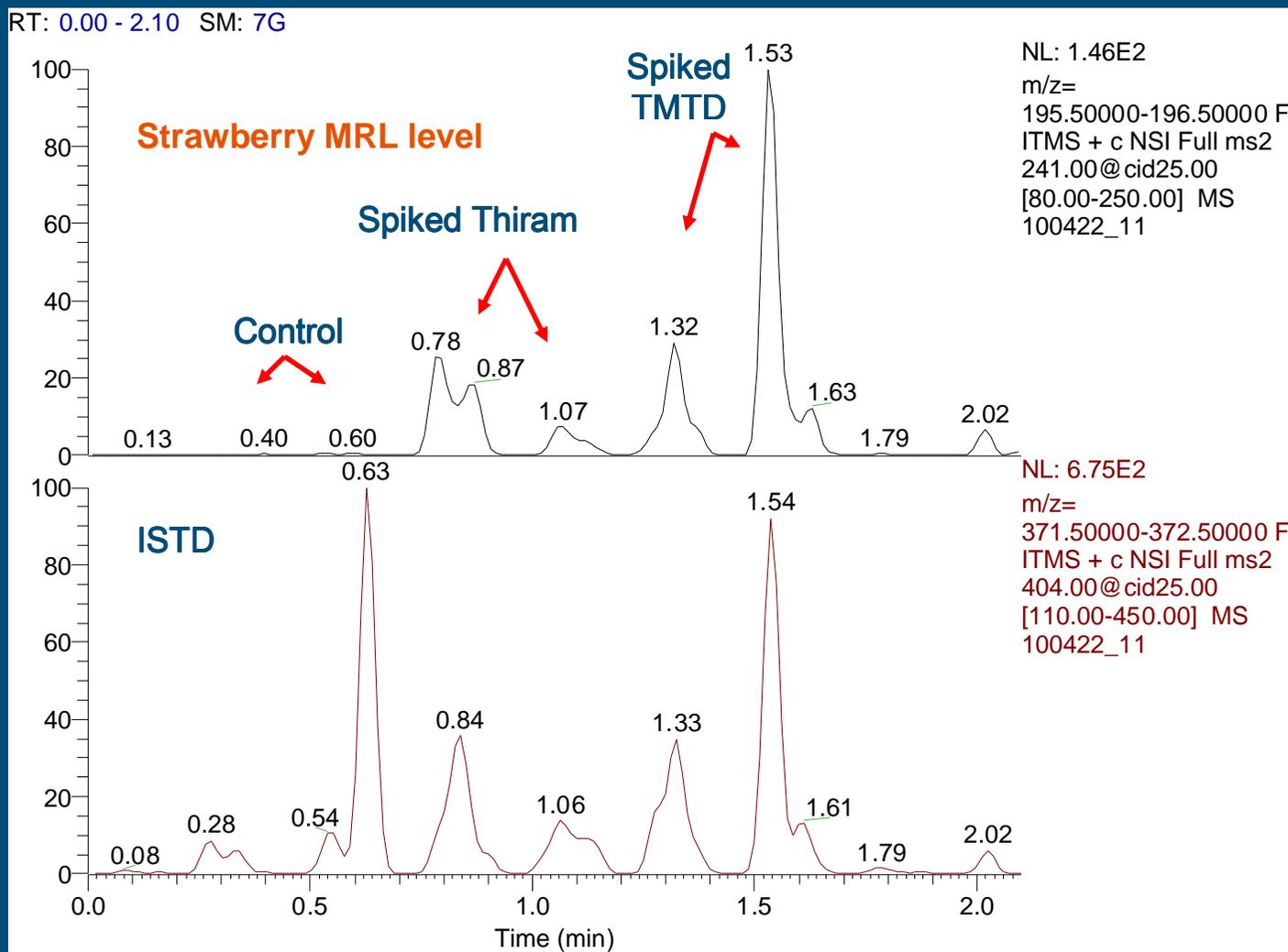
Vegetables/fruits spiked with thiram std and commercial PPP (TMTD)



1.5  $\mu$ l strawberry extracts

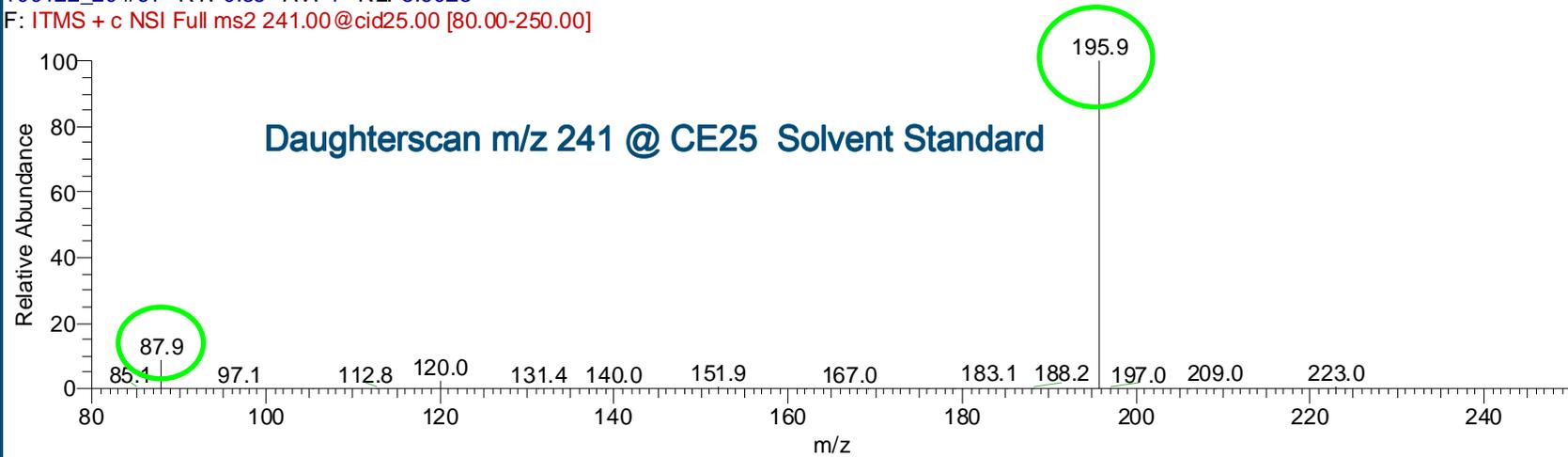
Ref. std thiram

# Results

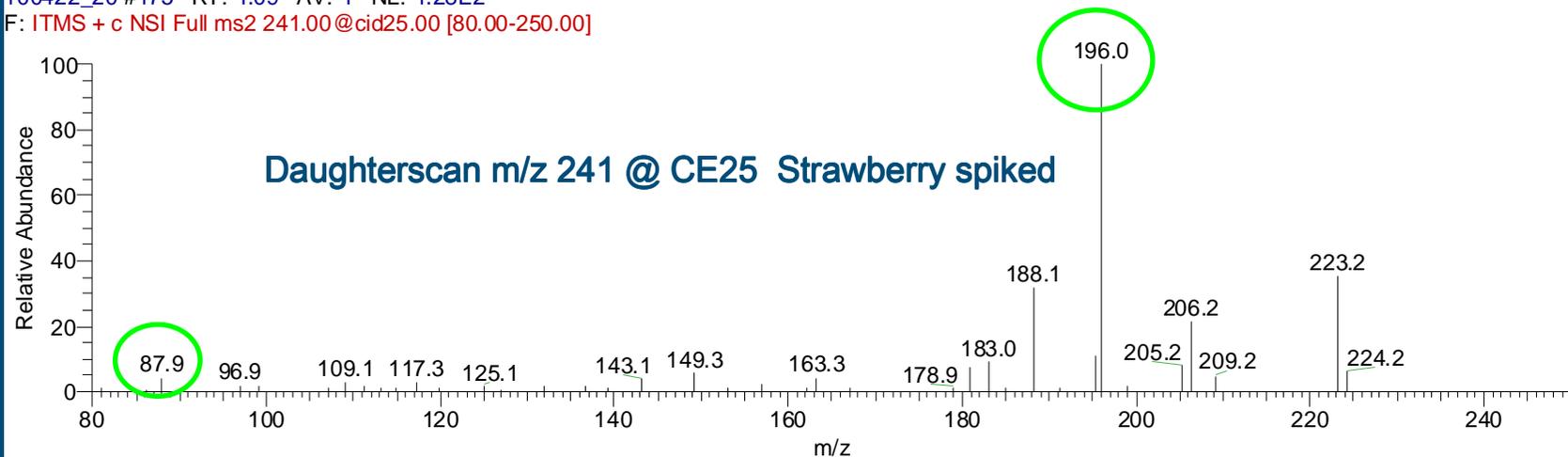


# Results

100422\_20 #61 RT: 0.38 AV: 1 NL: 8.60E3  
F: ITMS + c NSI Full ms2 241.00@cid25.00 [80.00-250.00]



100422\_20 #175 RT: 1.09 AV: 1 NL: 1.28E2  
F: ITMS + c NSI Full ms2 241.00@cid25.00 [80.00-250.00]



# Results

Recovery and repeatability (matrix-matched calibration)

Pears spiked (10 mg/kg)

	thiram	ISTD	ratio
1	3333	5568	0.60
2	9398	7561	1.24
3	6124	5759	1.06
4	6552	6623	0.99
5	3414	5054	0.68
6	5117	5490	0.93
average	5656	6009	0.92
RSD%	40	15	<b>26</b>
av. Rec.%			<b>85</b>

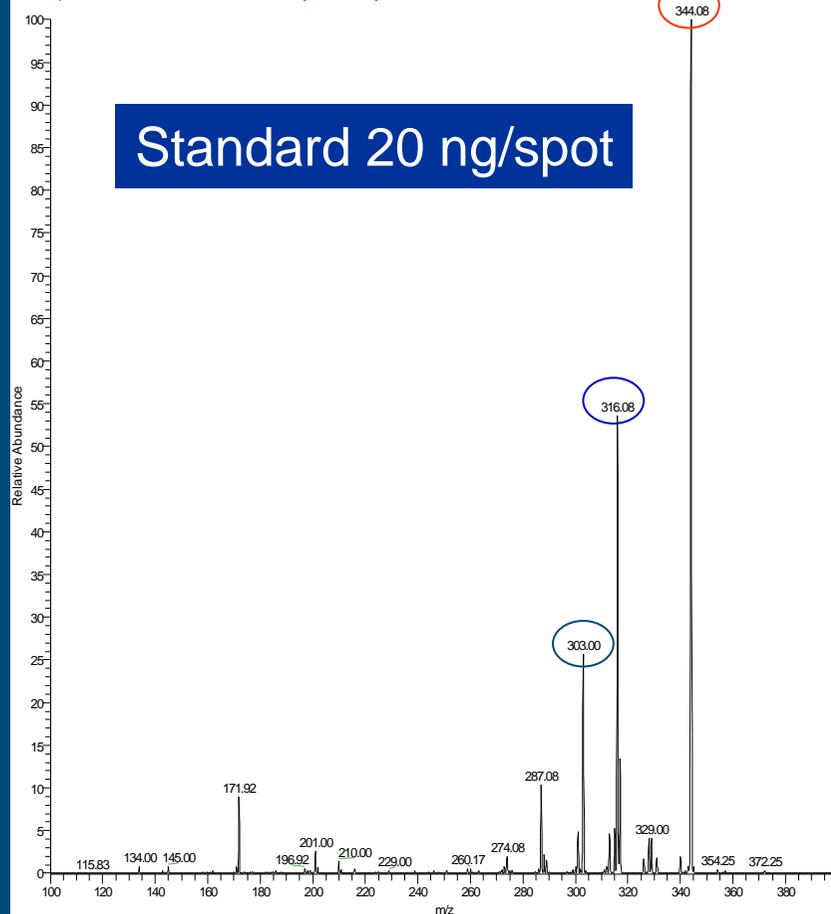
# Strobilurin fungicides (Biocop)



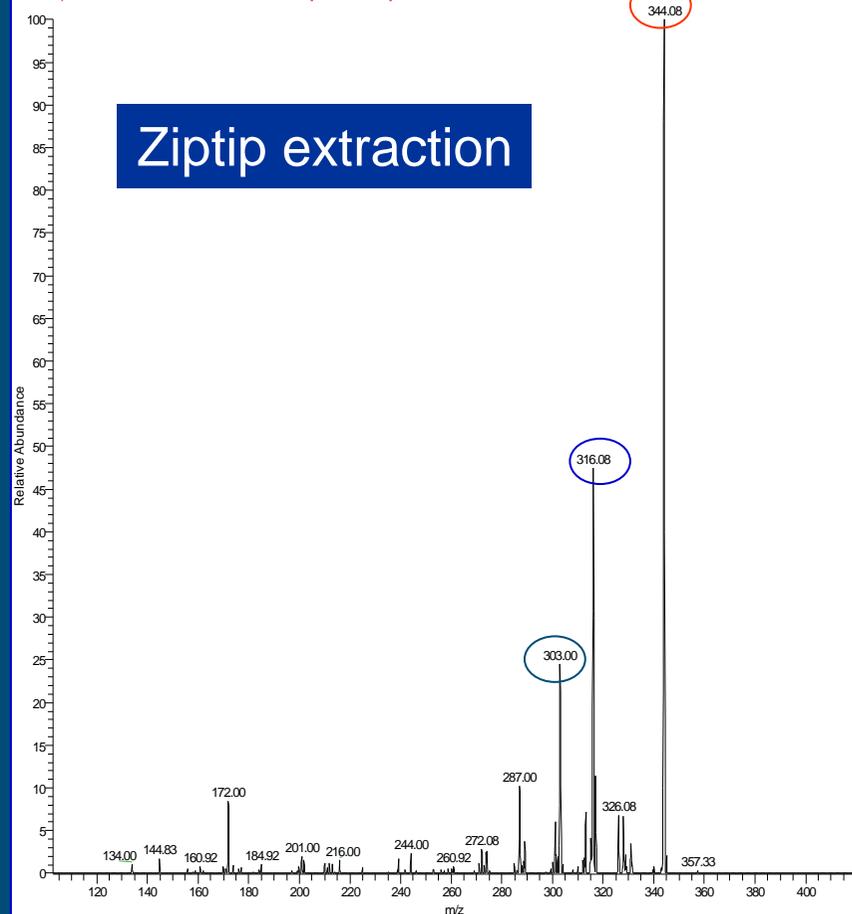
# Azoxystrobin MS/MS on m/z 404



071103\_hho\_13#13-24 RT: 0.34-0.66 AV: 12 NL: 2.51E1  
F: ITMS+p ESI Full ms3 404.00@cid30.00 372.00@cid30.00 [100.00-500.00]



071102\_hho\_20#239 RT: 7.13 AV: 1 NL: 1.14  
F: ITMS+p ESI Full ms3 404.00@cid35.00 372.00@cid35.00 [100.00-1250.00]



# Strobilurins: Reactive DESI

<i>strobilurin</i>	DESI from PTFE	Reactive DESI silver cationization
Metominostrobin "Z"	+	+
Metominostrobin "E"	+	+
Fenamidone	+	+/-
Kresoxim-methyl	-	+
Dimoxystrobin	+/-	+
Epoxiconazole	+	+/-
Famoxadone	-	+/-
Pyraclostrobin	-	+
Orysastrobin	-	+
Azoxystrobin	+	+
Trifloxystrobin	-	+
Fluacrypyrim	-	+
Fluoxastrobin	+	+
Picoxystrobin	+	+

DESI: ACN/water/FA (80:20:0.1)

reactive DESI: ACN/water (75:25) + 5 mg/ml AgCF<sub>3</sub>COOH flow 2.5 ul.min<sup>-1</sup>

# Pesticides in/on vegetables/fruits

Optimization of spray solvent

Compound	Limit of detection / ng ml <sup>-1</sup>					
	MeOH: H <sub>2</sub> O 50:50 <sup>a</sup>	MeOH: H <sub>2</sub> O 80:20 <sup>a</sup>	AcN: H <sub>2</sub> O 50:50 <sup>a</sup>	AcN: H <sub>2</sub> O 80:20 <sup>a</sup>	AcN: H <sub>2</sub> O 95:5 <sup>a</sup>	AcN: H <sub>2</sub> O 80:20 (1% FA <sup>b</sup> ) <sup>a</sup>
Ametryn	30	10	5	2.5	5	2.5
Amitraz	65	150	30	30	30	30
Atrazine	120	200	100	80	35	35
Azoxystrobin	16.5	35	25	25	65	16.5
Bitertanol	1000	400	250	300	500	250
Buprofezin	35	65	10	35	100	10
Imazalil	16.5	35	20	10	25	10
Imazalil metabolite	330	500	16.5	16.5	35	16.5
Isofenphos- methyl	200	200	130	35	200	35
Malathion	200	330	330	35	100	35
Nitenpyram	500	2000	50	100	100	50
Prochloraz	200	1650	150	150	250	150
Spinosad	30	35	130	35	330	30
Terbutylazine	150	250	130	130	65	65
Thiabendazole	165	400	330	100	50	50
Thiacloprid	300	1000	330	225	300	225

<sup>a</sup> All the experiments were performed using 3µl of solvent standards deposited on PTFE surfaces

<sup>b</sup>: FA: formic acid

# DESI parameters garcia et al

Nitrogen sheath gas	150 psi (10 bar)
Incident angle	55°
Spray tip-to-surface distance	5 mm
Surface to MS-inlet distance	1.5 mm
Scattering angle	ca. 10° to surface
DESI spray	acetonitrile/water 8:2+1% FA
Flow	5 µl/min
Scanning of the surface	?
Heated capillary MS (LTQ)	200°C
Substrate	PTFE

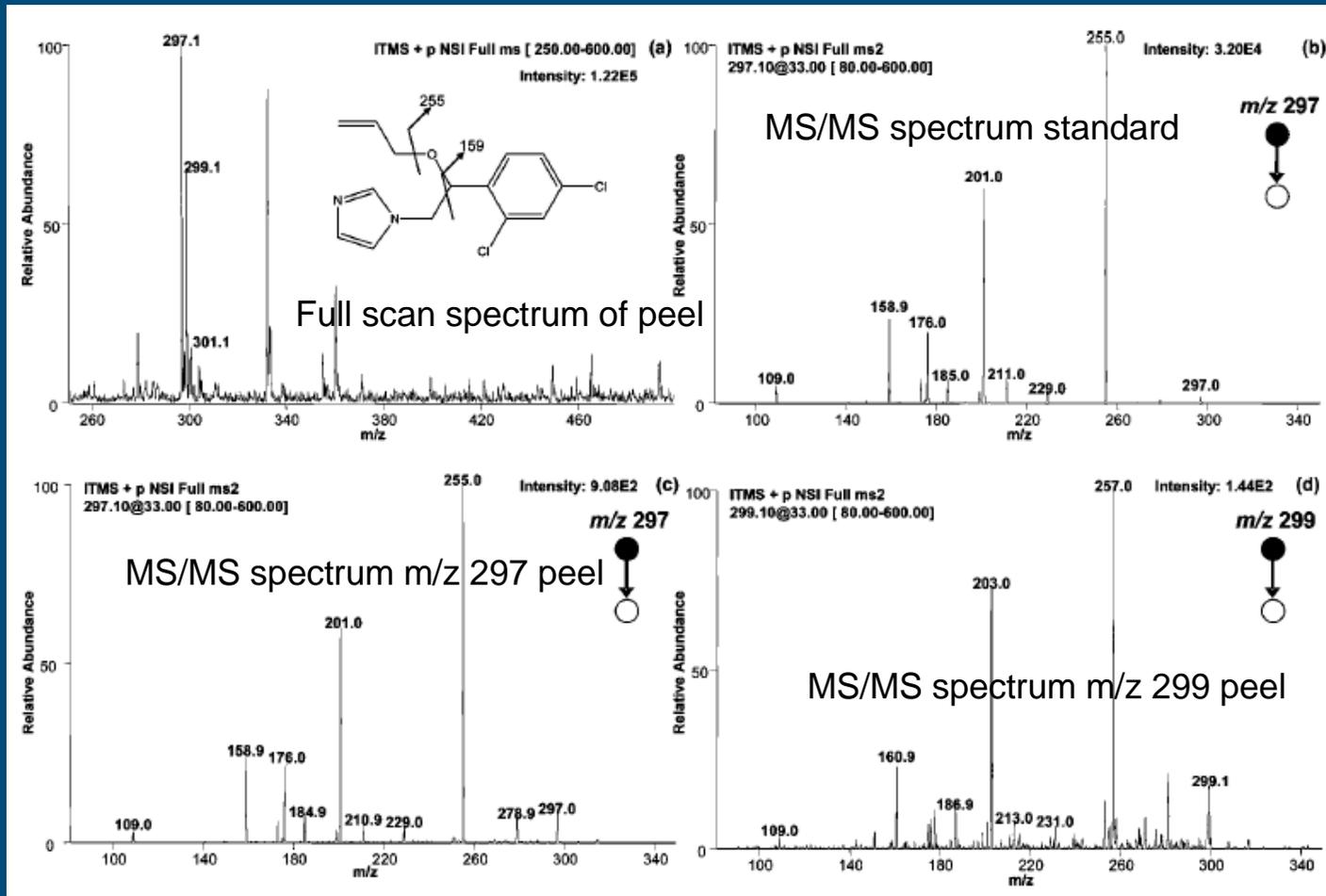
# Pesticides in/on vegetables/fruits

compound	limits of detection (LOD)			
	LOD ( $\mu\text{g L}^{-1}$ )		LOD ( $\mu\text{g kg}^{-1}$ )	
	DESI-MS (optimized solvent) <sup>c</sup>	DESI-MS/MS (solvent) <sup>d</sup>	DESI-MS/MS (tomato) <sup>e</sup>	DESI-MS/MS (orange) <sup>f</sup>
ametryn	2.7	0.1	1.0	1.5
amitraz	30	3.0	60	80
atrazine	30	0.16	1.5	1.5
azoxystrobin	15	2.0	18	18
bitertanol	265	6.5	60	80
buprofezin	10	2.0	80	80
imazalil	10	0.5	5.0	5.0
imazalil metabolite	17	5.0	90	90
isofenphos-methyl	35	3.5	80	60
malathion	35	3.0	30	40
nitenpyram	50	3.5	70	70
prochloraz	150	0.7	10	10
spinosad	30	8.0	80	80
terbuthylazine	65	0.16	2.0	2.0
thiabendazole	50	3.3	45	50
thiacloprid	225	5.0	50	60

QuEChERS w/o buffer + PSA clean up; extract diluted 1:3 in acetonitrile

# Pesticides in/on vegetables/fruits

Direct detection of imazalil from lemon peel



# Pesticides in/on vegetables/fruits

**Table 4. Quantitation of Imazalil Residues in Citrus Fruits by DESI-MS/MS and Comparison with the LC-MS Reference Method**

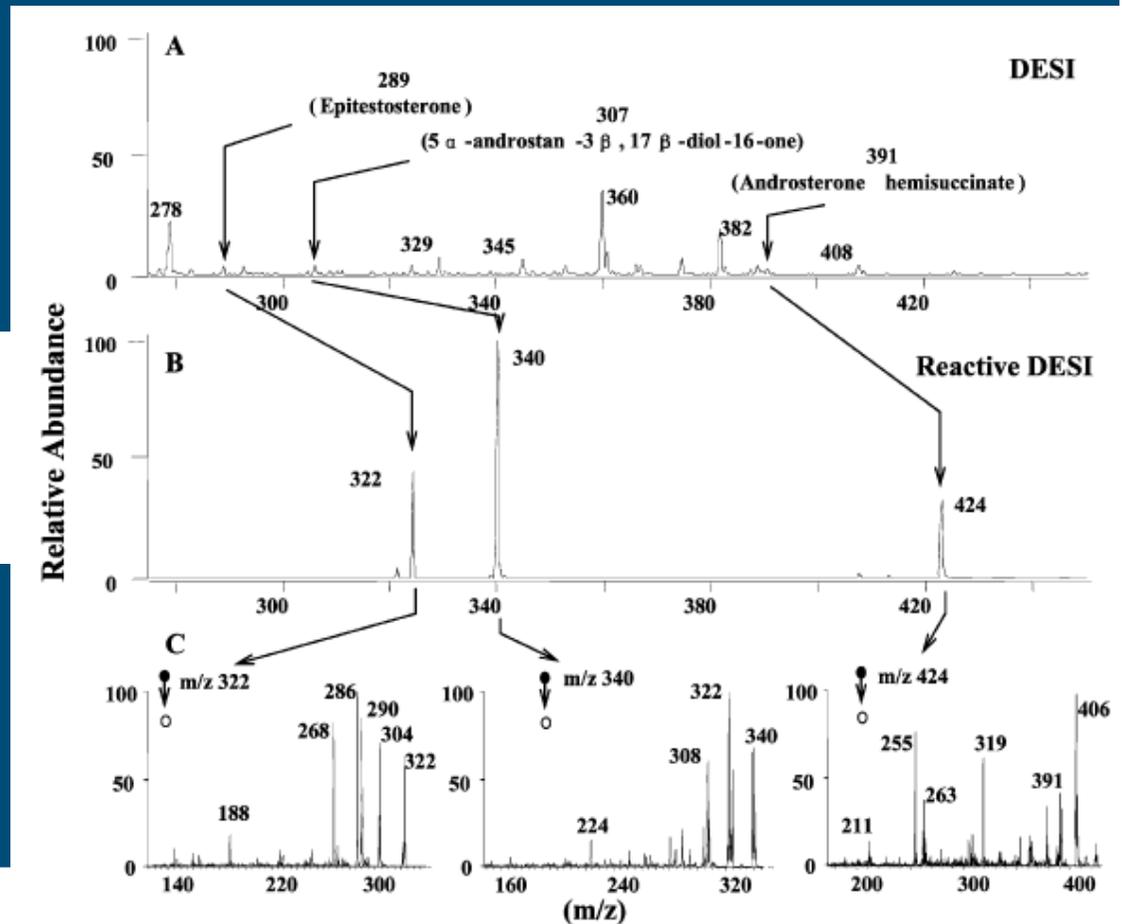
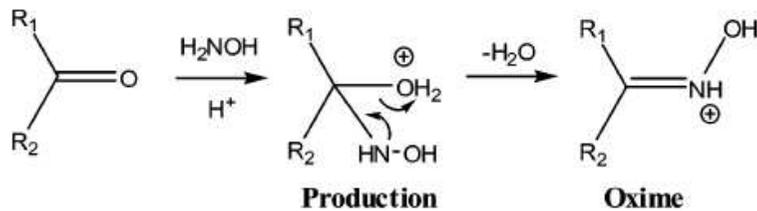
no.	sample	imazalil concentration ( $\mu\text{g g}^{-1}$ )		
		LC-MS ( $\mu\text{g g}^{-1}$ )	DESI-MS ( $\mu\text{g g}^{-1}$ )	RSD (%) ( $n = 5$ )
1	orange	0.19	0.16	12.8
2	lemon	0.35	0.38	11.2
3	lemon	0.44	0.48	9.6
4	orange	0.67	0.58	8.1
5	grapefruit	0.33	0.30	12.1
6	orange	0.15	0.12	14.5

using deuterated imazalil as ISTD

# Reactive DESI analysis on steroids

Steroids in urine  
SPME enrichment  
Direct detection from fiber

Spray 3 $\mu$ l/min:  
MeOH/water 1/1 0.05% HAc,  
5% hydroxylamine

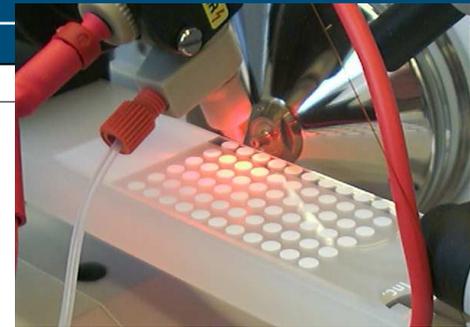
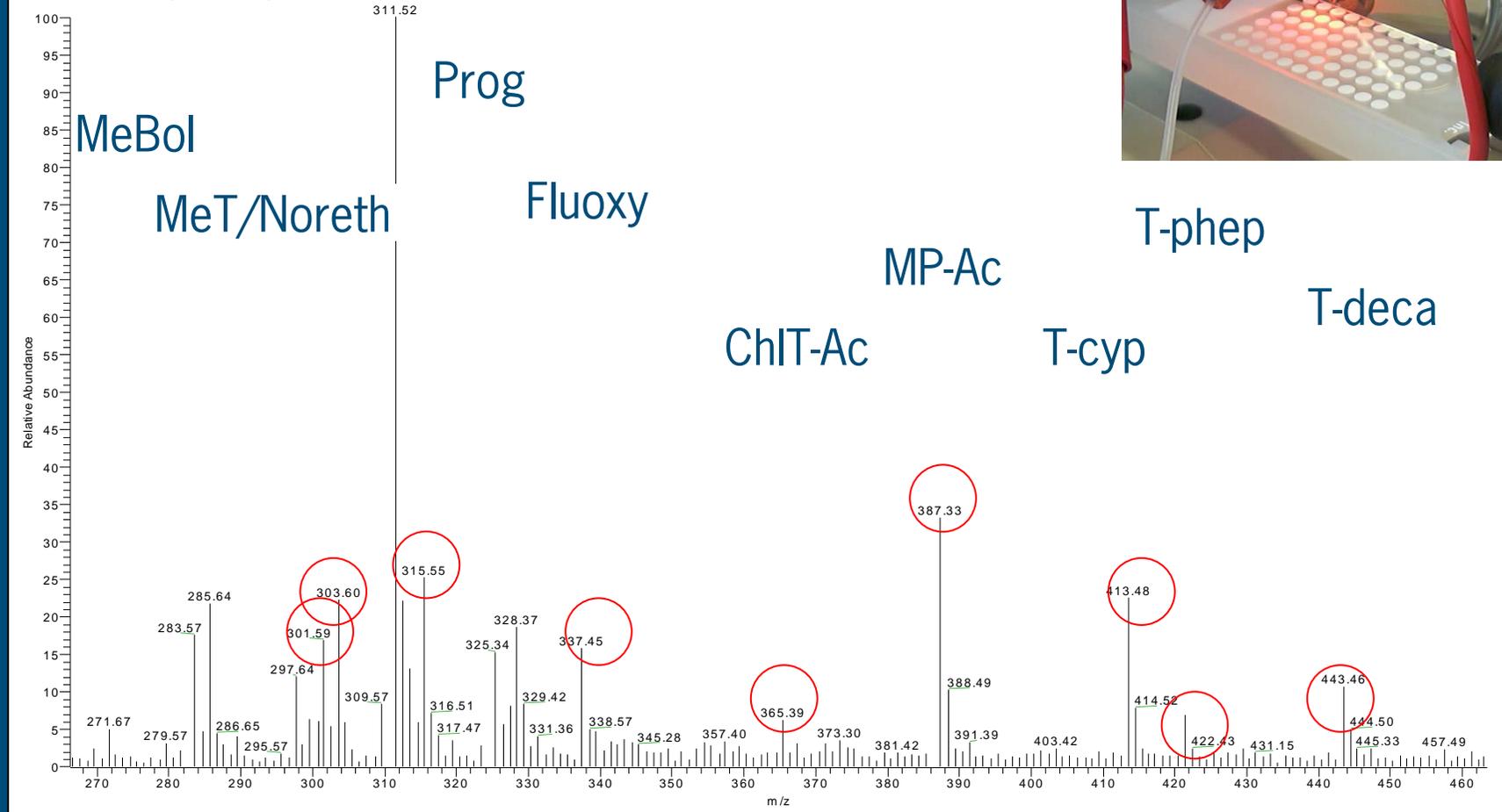


Huang et al, Anal. Chem. 79 (2007) 8327-8332

# Steroid cocktail

O:\Data\_DESIMS\...070912\070912\_hho\_\_40 9/12/2007 12:45:58 PM rik 167469  
2.5 µl/min MeOH/0.2 % FA 1/1 inj. vol.: 0.000000

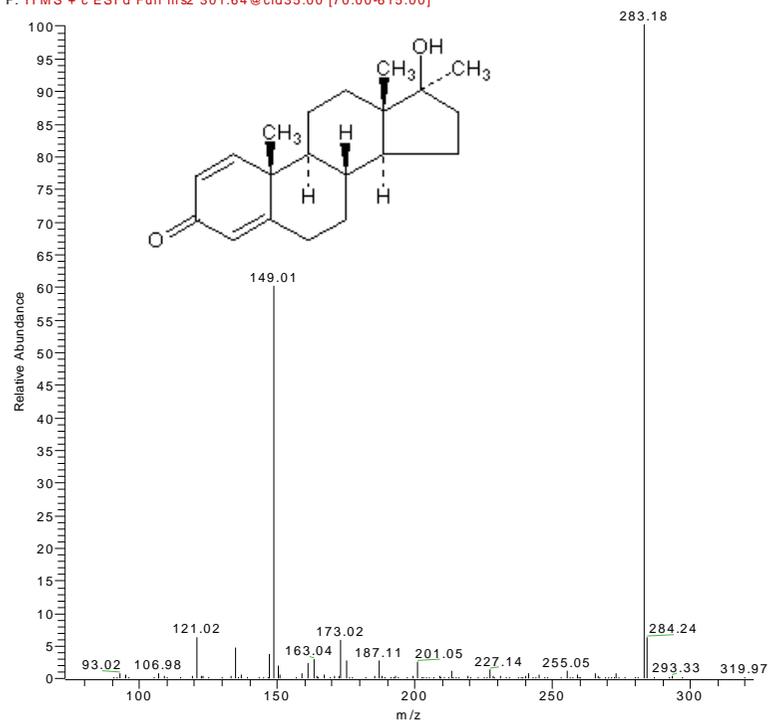
070912\_hho\_\_40 #31 RT: 0.08 AV: 1 NL: 5.14E3  
T: ITMS + c ESI Full ms [50.00-1250.00]



# Steroid cocktail

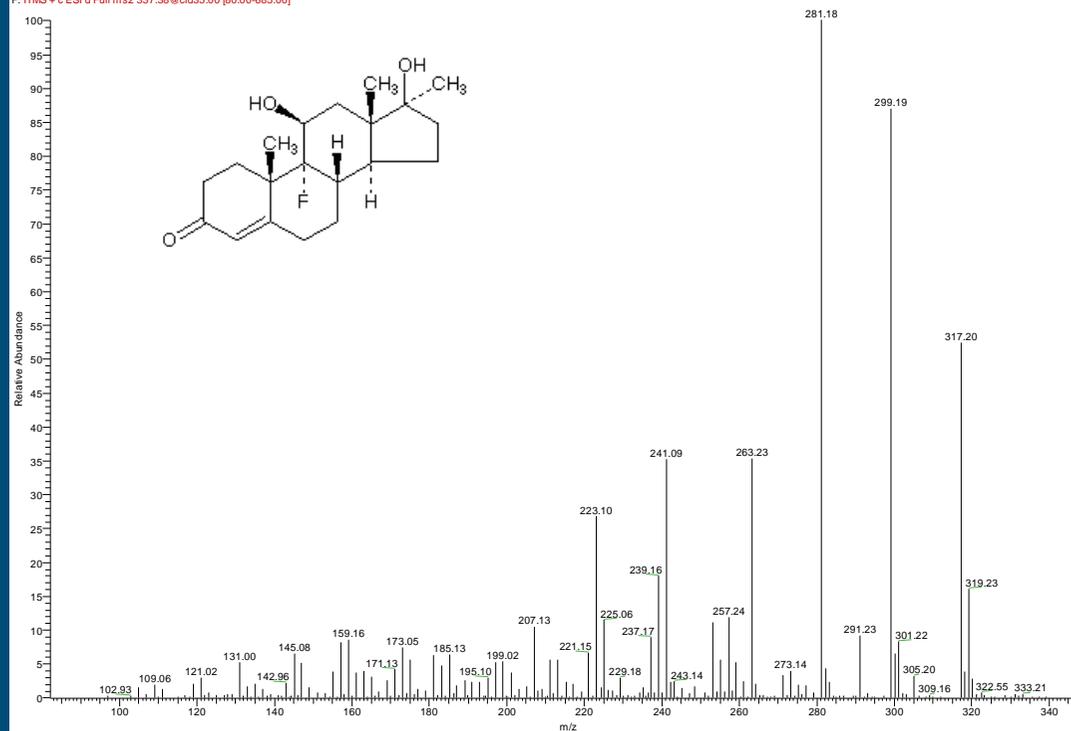
## Methylboldenone MS/MS m/z 301

070912\_hho\_40 #53 RT: 0.14 AV: 1 NL: 1.26E3  
F: ITMS + c ESI d Full ms2 301.64 @cid35.00 [70.00-615.00]



## Fluoxymesterone MS/MS on m/z 337

070912\_hho\_40 #2-35 RT: 0.03-0.08 AV: 3 NL: 2.81E2  
F: ITMS + c ESI d Full ms2 337.38 @cid35.00 [80.00-685.00]

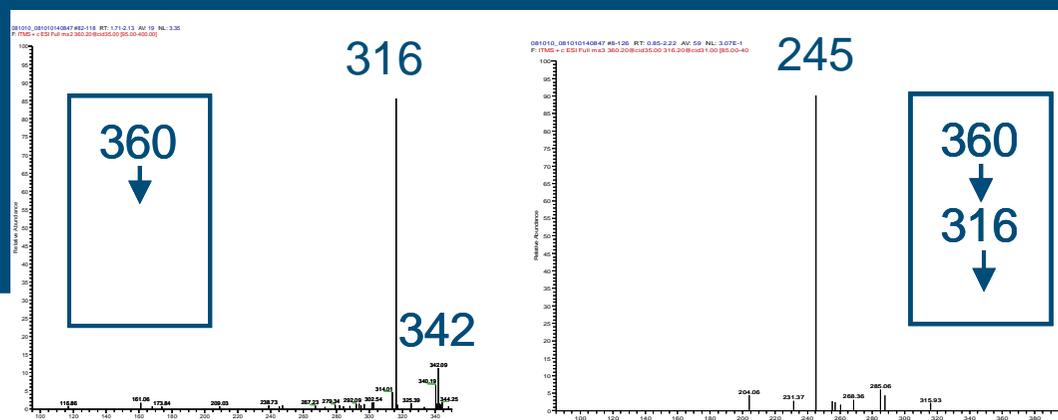
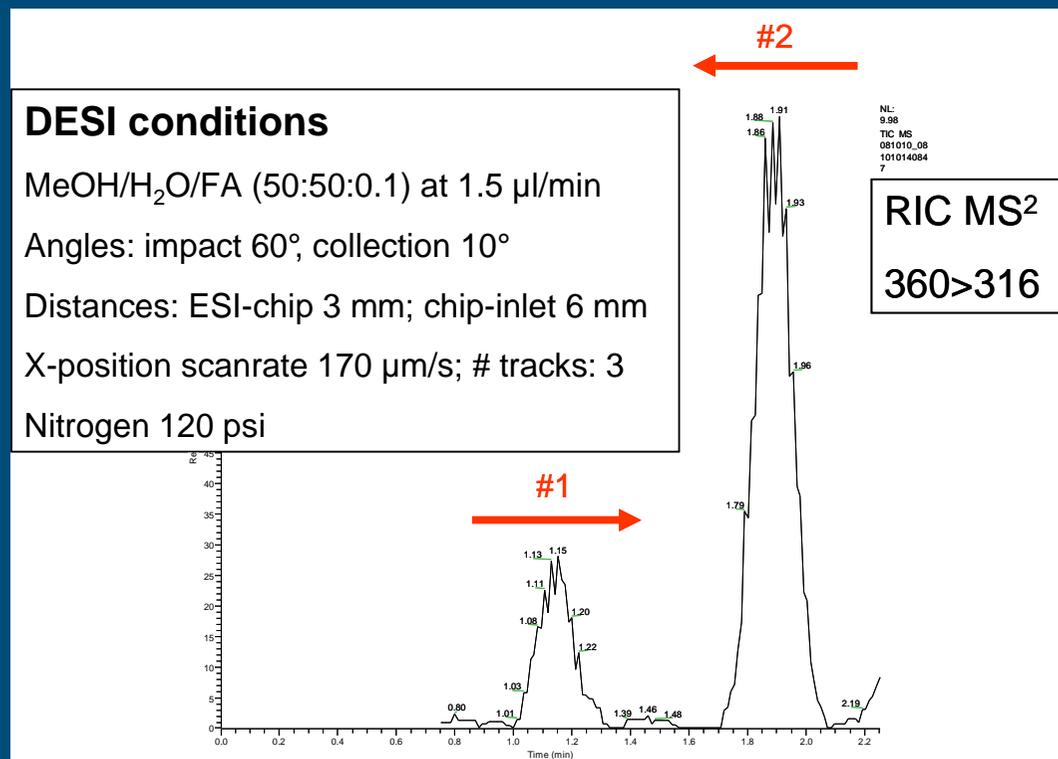


# Direct detection of IA captured fluoroquinolone

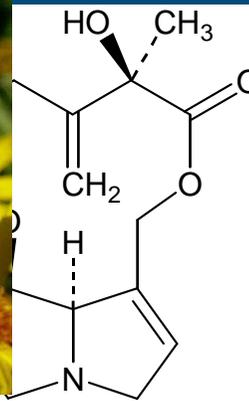
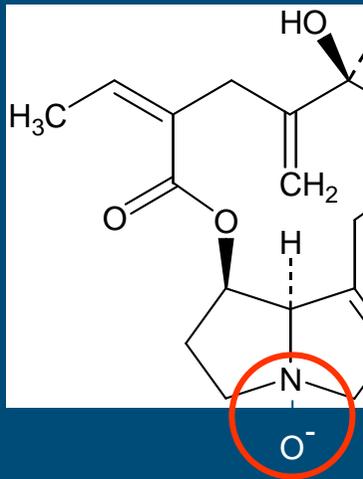
SPR biosensor chip  
Immunoaffinity bound enrofloxacin

## DESI conditions

MeOH/H<sub>2</sub>O/FA (50:50:0.1) at 1.5 µl/min  
Angles: impact 60°, collection 10°  
Distances: ESI-chip 3 mm; chip-inlet 6 mm  
X-position scanrate 170 µm/s; # tracks: 3  
Nitrogen 120 psi



# Alkaloids on *Senecio jacobaea* L. Stinking Willie



Senecionine N-oxide

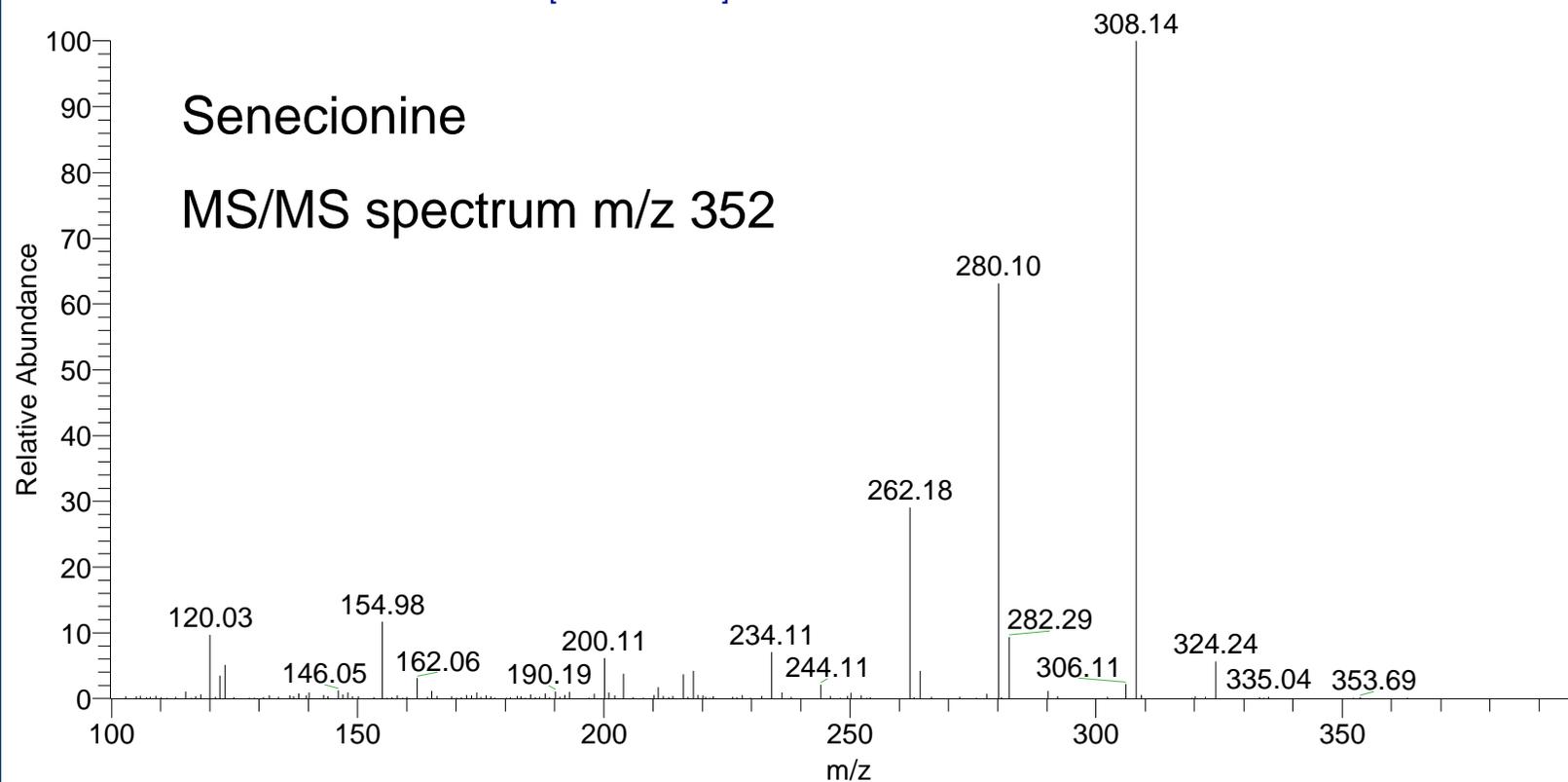
Unable to passively permeate membranes  
Nontoxic  
Metabolically safe

Senecionine

Able to passively permeate membranes  
Toxic after bioactivation  
Metabolically unsafe

# Alkaloids on *Senecio jacobaea* L. Stinking Willie

071003\_hho\_29 #16 RT: 0.38 AV: 1 NL: 6.49E2  
T: ITMS + c ESI Full ms2 352.00@cid35.00 [95.00-450.00]



# Summary

DESI a useful option for food contaminant analysis/control  
Lots of research in progress, many papers appearing  
Like other ambient techniques, pro's and con's

## Features:

- Direct detection from substrates, TLC, sensor chips, sample surface
- Reactive DESI
- Spatial resolution
- Rapid detection/high throughput of sample(extracts)

Quantitative determination possible with use of internal standard

## Limitations:

- Sensitivity (matrix suppression)
- Reproducibility
- MS<sup>n</sup> or high res required
- residues => targeted detection

# Outlook

## In lab analysis:

- Further improvement robustness
- Rapid straightforward and selective extraction
- More applications
- Combi sources to extent scope

ASAP+DESI (Lloyd et al, Anal. Chem. 81, 2009, 9158)

## Towards in-field:

- optimization of nongeometry dependent DESI source
- coupling of DESI with portable MS instruments
- .....

# Too much, too little, too fast, not clear?

## DESORPTION ELECTROSPRAY IONIZATION MASS SPECTROMETRY IN THE ANALYSIS OF CHEMICAL FOOD CONTAMINANTS

M.W.F. Nielen<sup>1,2\*</sup>, H. Hooijerink<sup>1</sup>, P. Zomer<sup>1</sup>, J.G.J. Mol<sup>1</sup>,

<sup>1</sup>, RIKILT Institute of Food Safety, P.O. Box 230, 6700 AE Wageningen, The Netherlands

<sup>2</sup>, Wageningen University, Laboratory of Organic Chemistry, Dreijenplein 8, 6703 HB

Wageningen, The Netherlands. E-mail: [michel.nielen@wur.nl](mailto:michel.nielen@wur.nl)

Submitted to Trends in Analytical Chemistry (March 2010)

# Thank you for your attention

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FP7-211326-CP "CONFIDENCE (Contaminants in Food and Feed: Inexpensive Detection for Control of Exposure)" coordinated by RIKILT – Institute of Food Safety (Wageningen, The Netherlands)

© Wageningen UR



# DESI, spray video



DESI spray on surface video clip.wmv

Venter, Nefliu, Cooks, Trends in Anal. Chem. 27 (2008) 284-290

# DESI, Alternative geometries

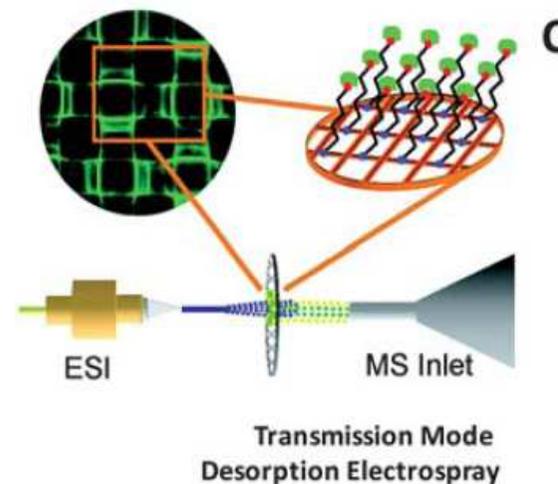
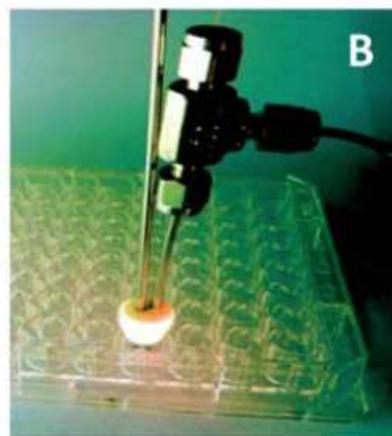
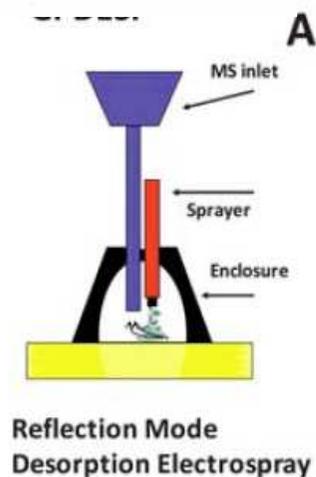
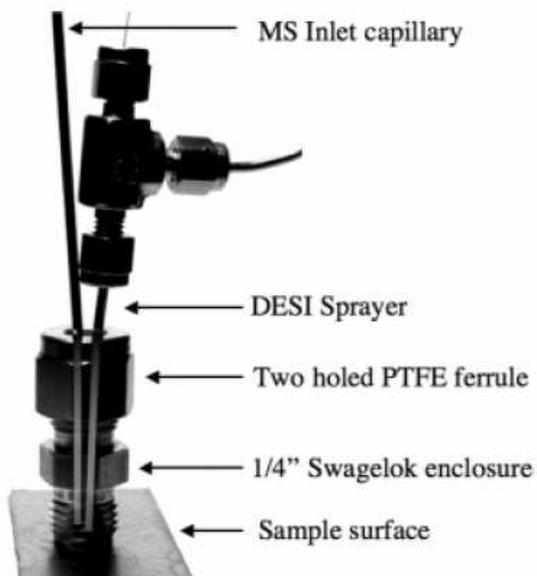


Fig. 4 Two simplified geometries for DESI. (A) and (B) Reflection mode (geometry independent) GI-DESI and its use in high-throughput analysis of metabolites in a bacterial matrix on a 96-well plate.<sup>24</sup> (C) Transmission mode (TM-DESI).<sup>27</sup>

# DESI, instrumentation

